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## MOLECULAR IDENTIFICATION OF BACTERIA OF GENUS STREPTOCOCCUS AND RELATED GENERA

The present invention pertains to the area of diagnosis. More precisely, the invention concerns a method for the molecular identification of bacteria of genus Streptococcus and related genera Enterococcus, Gemella, Abiotrophia and Granulicatella using detection and/or amplifying and sequencing techniques with probes or oligonucleotide primers applied to strains of these bacterial genera.

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10 Bacteria of the Streptococcus genus and of four related genera: Enterococcus, Gemella, Abiotrophia and Granulicatella, are Gram-positive and catalase-negative spherical bacteria of which more than around forty species are presently known. Bacteria of the genus Lactococcus, previously classified among 15 the streptococci as Group N Streptococcus, do not come within scope of this invention on account of their occurrence in human pathology, and because they can be easily distinguished from streptococci through their growth at +10°C. Genus Streptococcus officially comprises 55 species. Genus 20 Gemella comprises 6 species, genus Abiotrophia comprises 1 species, genus Granulicatella comprises 3 species, and genus Enterococcus comprises 24 species [www.springerny.com/bergeysoutline/main.htm]. These species are easily and frequently cultured from environmental samples, veterinary 25 clinical specimens and human clinical specimens [Ruoff Kl. (1999) in Manual of Clinical Microbiology, pp. 283-296, ASM Press]. In man, different species of the Streptococcus genus are responsible for community infections which may be severe

due the invasive nature of the streptococci consideration or through the production of possibly serious toxins with clinical signs distant from the site of infection. For example, Streptococcus pyogenes (Group A Streptococcus) is 5 responsible for throat infections and post-streptococcal syndromes including rheumatic fever during which damage to the heart valves through an inflammatory process is responsible for possibly fatal heart valve disease. Also, several species of genus Streptococcus, in particular Group A, Group C and 10 Group C Streptococci are responsible for life-threatening invasive infections, myositis in particular, i.e. degenerative changes to skin, subcutaneous and muscle tissue as has been described for some years. Also, Streptococcus pneumoniae (pneumococcus) for example causes pneumonia, meningitis and 15 septicaemia. Bacteria of the genera Streptococcus, Enterococcus, Abiotrophia and Granulicatella Gemella, can cause endocarditis i.e. infection of the heart valves in man, which come under life-threatening infectious diseases [Casalta et al., Journal Clinical Microbiology, 2002, 40: 20 some species of the genera under consideration 1847]. Also, can cause nosocomial infections, for example Streptococcus bacteria are responsible for subsequent to digestive endoscopy investigation. In addition, the of genus Enterococcus can cause nosocomial 25 urinary infections after prophylactic antibiotic therapy with cephalosporins against which they are naturally resistant. These bacterial species also raise the problem of their increasing resistance to antibiotics, the resistance Streptococcus pneumoniae to penicillin G [Garav J. 30 Infect. Dis. 2002, 2: 404-415] and the resistance Enterococcus spp. to vancomycin [Gold H.S., Clin. Infect. Dis. 2001, 33: 210-219; Bonten M.J. et al. Lancet Infect. 2001, 1: 314-325].

These different bacterial species raise the problem of their detection in human pathological specimens and of their identification when isolated from such samples. Conventional detection methods rely on the evidencing of Gram-positive cocciform bacteria on direct examination of the pathological specimen. Ιt is known, however, that this microscopic detection of bacteria of the genus Streptococcus and related genera in clinical specimens has a sensitivity threshold of 104 CFU/ml. It is therefore fully possible that a pathological specimen in man or animal contains one of the species under consideration which is not detected by direct microscopic examination of this pathological specimen. In addition, even though their structure is of Gram-positive bacterial type, they may give a false Gram-negative result after Gram staining of the pathological sample and give rise to erroneous or inconclusive identification. This is particularly frequent in bacteria of genus Gemella. In man, this is especially the case in anatomopathological and bacteriological investigation of the heart valves when diagnosing endocarditis.

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When a bacterium of one of the species of the genera consideration is isolated in the laboratory, conventional phenotype identification methods are the most commonly used to identify bacteria of species belonging to Streptococcus and related genera, and identification kits and automated analysers have developed to assist phenotype identification of bacteria of genus Streptococcus and related genera. In this respect, the extent of identification in routine practice is variable. In particular, one of the tests used for identifying Streptococci and bacteria of related genera is the detection of haemolytic reaction, i.e. the destruction by the bacterium of red blood cells contained in a blood agar. However, this haemolytic reaction can be inhibited by the presence of oxygen

or by the presence of a peroxide when Streptococci bacteria cultured in the presence of a high carbon dioxide concentration. Moreover, it is recognized that there exists a certain extent of subjectivity in assessing haemolysis by 5 colonies of Streptococci and hence inter-operator variability which is detrimental to the quality of identification of these bacteria. For alpha-haemolytic streptococci, a second test is the optochin sensitivity test which enables identification of Streptococcus pneumoniae which is sensitive to this compound. 10 However, strains of Streptococcus pneumoniae resistant to optochin have been reported [Lund E. Acta Patho. Microbiol. Immunol. Scand. 1959, 47, 308-315]. A final phenotype test is which may also give false positive results in particular for streptococci in serogroup D on account of cross 15 antigenicity between group D streptococci, Enterococcus and Pediococcus.

Several molecular systems have been developed to identify some serogroups or some species of genus Streptococcus, in particular for group A streptococci (Streptococcus pyogenes, 20 Streptococcus aginosus, Streptococcus constellatus, Streptococcus intermedius) and group (Streptococcus В agalactiae) [Daly J.A. et al. J. Clin. Microbiol. 1991, 29:80-82; Heelan J.S. et al., Diagn. Microbiol. Infect. Dis. 1996, 65-69] and for Streptococcus pneumoniae [Denys G.A. & Carrey R.B., J. Clin. Microbiol. 1992, 30: 2725 - 2727] by 25 hybridisation of specific probes targeting the gene encoding the 16S ribosomal RNA. Also, different systems based on PCR amplification of genes coding for toxins or virulence factors have been developed to discriminate Streptococcus pneumoniae 30 from among  $\alpha$ -haemolytic Streptococci [Salo P. et al., J. Infect. Dis. 1995, 171: 479-482; Morrisson K. et al. J. Clin. 2000, Microbiol. 38, 434-437; Kaijalainen т. et J. Microbiol. Meth. 2002, 51: 111-118], and for the detection of

Streptococcus agalactiae [Mawn J.A. et al. J. Clin. Pathol. 1993, 46: 633-636]. These different systems, however, only allow the identification of one or of a few species of genus Streptococcus.

5 An identification system for three species of streptococcus has been developed, based on amplification of the 16S-23S spacer [Forstman P. et al. Microbiology, 1997, 143, 3491-3500] but in this work identification was limited to of few species animal interest: Streptococcus 10 agalactiae, Streptococcus dysgalactiae and Streptococcus uberis. Also, at the present time it is essential for laboratories to have 2 separate molecular targets for the detection and identification of streptococci to overcome the risks of molecular contamination inherent in the use of a 15 single target.

Finally, no detection and identification system for Streptococcus-related genera has been developed, and more particularly for bacteria of the genera Enterococcus, Gemella, Abiotrophia and Granulicatella.

The inventors have shown in the present invention that the rpoB gene forms a genetic marker which can be used for the detection and specific identification of the bacterium of each species in genus Streptococcus and in 4 related genera: Enterococcus, Gemella, Abiotrophia and Granulicatella.

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Although this gene has previously been shown to have use as a tool in bacterial identification of different bacterial no publication mentions its use for identifying bacteria of genus Streptococcus and the four related genera, and the advantage of this gene's sequence for the identification of the said bacteria has in no way suggested. On the contrary, a few partial sequences of the rpoB gene in a few species, available in GenBank, slight heterogeneity placing in doubt the advantage of this gene as an identification tool for these bacteria. Finally, have developed a tool for the simultaneous inventors identification of four bacterial genera, requiring development of degenerate primers which could not be deduced from any of the rpoB sequences determined for each species.

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More particularly, the present invention concerns nucleic acid sequences specific to the genus or to each species of Streptococcus and related genera whose nucleotide sequence is derived from the rpoB gene of the said bacteria.

According to Lazcano et al. [J. Mol. Evol. (1988) 27: 365-376] the polymerase RNAs are divided into two groups as per their origin, one consisting of the RNA- or DNA-dependent viral polymerase RNAs and the other consisting of the DNAdependent polymerase RNAs of eukaryote or prokaryote origin (archaebacteria and eubacteria). The eubacterial DNA-dependent polymerase RNAs are characterized by a simple, conserved multimeric constitution denoted "core enzyme" represented by  $\alpha\beta\beta'$ , or "holoenzyme" represented by  $\alpha\beta\beta'\sigma$  [Yura and Ishihama, Ann. Rev. Genet. (1979) 13: 59-57].

Numerous studies have evidenced the functional within the multimeric enzymatic complex, of the  $\beta$  subunit of the eubacterial polymerase RNA. Archaebacterial and eukaryote polymerase RNAs have а more complex structure possibly reaching ten and even thirty subunits [Pühlet et al. Proc. 25 Natl. Acad. Sci. USA (1989) 86: 4569-4573].

The genes encoding the different  $\alpha\beta\beta'\sigma$  subunits of the DNA-dependent polymerase RNA in eubacteria, the genes rpoA, rpoB, rpoC and rpoD respectively, are classified in different groups comprising the genes coding for constituent proteins of ribosomal subunits or for involved enzymes in the replication and repair of the genome [Yura and Yshihma, Ann. Rev. Genet. (1979) 13: 59-97]. Some authors have shown that the sequences of the rpoB and rpoC genes could be used to construct phylogenetic trees [Rowland  $et\ al$ . Biochem. Soc. Trans. (1992) 21:40S] enabling separation of the different branches and sub-branches among the kingdoms of the living.

Before setting forth the invention in more detail, different terms used in the description and claims are defined below:

- By "nucleic acid extracted from bacteria" is meant either the total nucleic acid, or the genomic DNA, or the messenger RNAs, or the DNA obtained from reverse transcription of the messenger RNAs.

- A "nucleotide fragment" or an "oligonucleotide" synonymous terms designating a chain of nucleotide motifs characterized by an information sequence of the natural (or optionally modified) nucleic acids and able to hybridise, 15 like natural nucleic acids, with a complementary or substantially complementary nucleotide fragment predetermined conditions of high stringency. The chain may contain nucleotide motifs having a different structure to natural nucleic acids. Α nucleotide fragment (or 20 oligonucleotide) may for example contain up 100 nucleotide motifs. It generally contains at least 8, and in at least 12 nucleotide particular motifs, particularly 18 to 35, and may be obtained from a natural nucleic acid molecule and/or by genetic recombination and/or 25 by chemical synthesis.
- A nucleotide motif is derived from a monomer which may be a natural nucleotide of a nucleic acid whose constituent elements are a sugar, a phosphate group and a nitrogenous base chosen from among adenine (A), guanine (G), uracil (U), cytosine (C), thymine (T); or else the monomer is a nucleotide modified in at least one of the three preceding constituent elements; as an example, modification may occur either at the bases, with modified bases such as inosine

which can hybridise with any base A,T,U,C or G, methyl-5-deoxycytidine, deoxyuridine, dimethylamino-5-deoxyuridine or any other modified base able to hybridise, or at the sugar, for example the replacement of at least one deoxyribose by a polyamide (Nielsen PE et al., Science (1991) 254: 1497-1500], or at the phosphate group, for example through replacement by esters chosen from among diphosphates, alkylphosphonates and phosphorothioates.

- By "hybridisation" is meant the process during which, under 10 suitable conditions, two nucleotide fragments sufficiently complementary sequences are able together by stable, specific hydrogen bonds to form a double Hybridisation conditions are determined "stringency" i.e. the strictness of operating conditions. 15 Hybridisation is more specific the higher the stringency. Stringency depends in particular upon the base composition of a probe/target duplex and on the extent of mismatch between two nucleic acids. Stringency may also be related to parameters of the hybridisation reaction, such as the 20 concentration and type of ion species present solution, type hybridisation the and concentration denaturing agents and/or the temperature of hybridisation. The stringency of the conditions in which a hybridisation reaction must be conducted depends in particular upon the 25 probes used. All this data is well known and the suitable conditions may possibly be determined in each case by routine experiments. In general, depending upon the length of the probes used, the temperature for the hybridisation reaction between approximately 20 65°C, lies and 30 particular between 35 and 65°C in a saline solution at a concentration of around 0.8 to 1 M.
  - A "probe" is a nucleotide fragment having hybridisation specificity under determined conditions to form a

hybridisation complex with a nucleic acid having, in this case, a nucleotide sequence included either in a messenger RNA or in a DNA obtained by reverse transcription of said messenger RNA, the transcription product; a probe may be used for diagnosis purposes (capture and detection probes in particular) or for therapeutic purposes.

- A "capture probe" is a probe that is or may be immobilised on a solid support by any appropriate means, for example by covalency, adsorption, or direct synthesis on a solid. Examples of supports include microtitration wafers and DNA
- Examples of supports include microtitration wafers and DNA chips.

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- A "detection probe" is a probe labelled with a marking agent chosen for example from among radioactive isotopes, enzymes particular enzymes able to act on a chromogenous, 15 fluorigenous or luminescent substrate (in particular a peroxidase alkaline phosphatase), or an chromophorous chemical compounds, chromogenous, fluorigenous luminescent compounds, analogues of nucleotide bases and ligands such as biotin.
- 20 A "species probe" is a probe enabling the specific identification of the species of a bacterium.
  - A "genus probe" is a probe enabling the specific identification of the genus of a bacterium.
- A "primer" is a probe having 10 to 100 nucleotide motifs for example and having hybridisation specificity under determined conditions for enzymatic amplification reactions.
  - By "amplification reaction" is meant an enzymatic polymerisation reaction, for example in an amplification technique such as PCR, initiated by primer oligonucleotides and using a polymerase DNA.
  - By "sequencing reaction" is meant the obtaining of the sequence of a nucleic acid fragment or of a complete gene by means of an abortive polymerisation method using

oligonucleotide primers and said dideoxynucleotides [Sanger F, Coulson AR (1975), J. Mol. Biol. 94: 441] or multiple hybridisations with multiple probes fixed on a solid support such as used in DNA chips for example.

The sequences of the rpoB genes of the bacteria Streptococcus pneumoniae, Streptococcus pyogenes, Streptococcus mutans and Streptococcus agalactiae have been described in the literature.

The inventors have determined the complete sequences of 10 of other bacterial species of genes Streptococcus and related genera: Streptococcus anginosus and Streptococcus equinus, Abiotrophia defectiva, and a very large portion of the gene for Streptococcus mutans and Enterococcus faecalis. These species were chosen by the inventors 15 representing the main genetic groups determined on the basis of the study on 16S the gene in bacteria of Streptococcus and related genera, encompassing all the species currently described in this genus, so that the alignment of the rpoB sequences obtained in these species would most 20 probably encompass all the rpoB sequences of all the species of these bacterial genera, more precisely they are therefore the most divergent species from a phylogenetic viewpoint among all the species currently described in this genus, so that the alignment of the rpoB sequences obtained in these 25 would most probably from a phylogenetic viewpoint encompass all the rpoB sequences of all the species of this bacterial genus.

From these complete or almost complete sequences, and after numerous unsuccessful attempts such as reported in examples 1 and 2 below, the inventors have evidenced the following consensus and specific sequences SEQ ID n°6 and 7:

- SEQ ID N°6: 5'- AARYTNGGMCCTGAAGAAAT-3', and
- SEQ ID N°7: 5'- TGNARTTTRTCATCAACCATGTG-3'

in which:

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- N represents inosine or one of the 4 nucleotides A, T, C or  $\mathsf{G}$ ,
- R represents A or G,
- M represents A or C, and
  - Y represents C or T,

and the reverse sequences and complementary sequences.

The inventors have determined said sequences SEQ ID n°6 and 7 as being not only consensual between all the bacteria of genus *Streptococcus* and said 4 related genera, but also specific to the family of bacteria of genus *Streptococcus* and said 4 related genera.

At the position corresponding to a nucleotide N,Y,M or R in sequences SEQ ID n°6 and 7, variable nucleotides are found in the complementary target sequences in relation to the species of the bacterium under consideration, but all the other nucleotides are conserved in all the species of bacteria of genus *Streptococcus* and of said 4 related genera.

Sequences SEQ ID n°6 and 7 so defined are present in the rpoB genes of all bacteria of genus Streptococcus and said 4 related genera, and are specific to the bacteria of genus Streptococcus and said 4 related genera, and can therefore provide genus probes or amplification primers to detect any bacterium of genus Streptococcus and of said 4 related genera.

For this purpose, one subject of the present invention is therefore an oligonucleotide which comprises a sequence of at least 8, preferably at least 12, further preferably between 18 and 35 nucleotide motifs, of which at least one sequence of 8, preferably 12, further preferably 18 consecutive motifs is included in one of the following sequences SEQ ID n°6 and 7:

- SEQ ID N°6: 5'-AARYTNGGMCCTGAAGAAAT-3', and
- SEQ ID N°7: 5'-TGNARTTTRTCATCAACCATGTG-3'

in which:

- N represents inosine or one of the 4 nucleotides A, T,  $\mbox{\ensuremath{\text{C}}}$  or  $\mbox{\ensuremath{\text{G}}},$
- R represents A or G,
- M represents A or C, and
- 5 Y represents C or T

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and the reverse sequences and complementary sequences.

Another subject of the invention is a mixture of oligonucleotides characterized in that it consists of an equimolar mixture of oligonucleotides of the invention, all having a different sequence and all comprising a sequence included in SEQ ID  $n^{\circ}6$  or all comprising a sequence included in SEQ ID  $n^{\circ}7$ .

More particularly, a further subject of the invention is a mixture of said oligonucleotides, consisting of an equimolar mixture of 32 oligonucleotides of different sequences each comprising at least 15, preferably at least 18 consecutive nucleotide motifs included in the following sequence:

- SEQ ID n°6: 5' AARYTNGGMCCTGAAGAAAT-3' in which:

- R represents A or G,
  - Y represents C or T
  - M represents A or C, and
  - N represents A, T, C or G

and the reverse sequences and complementary sequences.

- A further subject of the invention is a mixture of said oligonucleotides consisting of an equimolar mixture of 8 oligonucleotides having different sequences and each comprising at least 15, preferably at least 18 consecutive nucleotide motifs included in the following sequence:
- 30 SEQ ID n°6: 5' AARYTNGGMCCTGAAGAAAT-3' in which:
  - R represents A or G,
  - Y represents C or T

- M represents A or C, and
- N represents inosine

and the reverse sequences and complementary sequences.

A further subject of the invention is a mixture of said oligonucleotides, consisting of an equimolar mixture of 16 oligonucleotides having different sequences and each comprising at least 15, preferably at least 21 consecutive nucleotide motifs included in the following sequence:

- SEQ ID n° 7: 5' TGNARTTTRTCATCAACCATGTG-3'
- 10 in which:

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- R represents A or G, and
- N represents A, T, C or G

and the reverse sequences and complementary sequences.

A further subject of the present invention is a mixture of said oligonucleotides, consisting of an equimolar mixture of 4 oligonucleotides having different sequences and each comprising at least 15, preferably at least 21 consecutive nucleotide motifs included in the following sequence:

- SEQ ID n° 7: 5'-TGNARTTTRTCATCAACCATGTG-3'
- 20 in which:
  - R represents A or G, and
  - N represents inosine,

and the reverse sequences and complementary sequences.

Said mixtures of oligonucleotides are able to hybridise with a complementary sequence included in the *rpoB* gene of all the bacteria of genus *Streptococcus* and said 4 related genera, and can therefore be used as a genus probe or as amplification primers for the detection or respectively the amplification of an *rpoB* gene fragment of said bacterium.

To prepare said equimolar mixture of oligonucleotides using oligonucleotide synthesis methods known to persons skilled in the art, an equimolar mixture is used of 4 or 2

nucleotides for the nucleotides corresponding to N or respectively K,N,R or Y, namely:

 an equimolar mixture of the 4 nucleotides A, T, C and G for the nucleotides corresponding to N in which N represents A, T, C or G, and

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- an equimolar mixture of the 2 nucleotides T and G for the nucleotides corresponding to K,
- an equimolar mixture of the 2 nucleotides A and C for the nucleotides corresponding to N,
- 10 an equimolar mixture of the 2 nucleotides A and G for the nucleotides corresponding to R, and
  - an equimolar mixture of the 2 nucleotides C and T for a nucleotide represented by Y.

Hence an equimolar mixture is obtained of 32  $(2^3x4)$  and 15  $16(2^2x4)$  nucleotides of different sequences for the 2 sequences SEQ ID n°6 and 7 respectively.

In said equimolar mixtures of oligonucleotides according to the invention, since "N" represents inosine which is able to hybridise with any base or an equimolar mixture of the 4 bases A, T, C, G, the sequences SEQ ID  $n^{\circ}$  6 and 7 are able to hybridise with the complementary sequence included in the rpoB gene of all bacteria of the Streptococcus genus and of the said 4 related genera.

In addition, these consensus sequences SEQ ID n°6 and n°7 flank hyper-variable sequences whose sequence is specific to each bacterium species of genus *Streptococcus*. These sequences flanked by SEQ ID n°6 and 7 may therefore be used as species probe for the bacteria of genus *Streptococcus* and said 4 related genera.

In addition, the sequences SEQ ID n°6 and 7 were determined as flanking an *rpoB* gene fragment comprising a zone whose variable length is approximately 720 bp and as

comprising the shortest sequences specific to each bacterium species of the *Streptococcus* genus and said 4 related genera.

The inventors were therefore able to evidence species probes for each of the examined 28 bacterial species of genus Streptococcus and said 4 related genera, corresponding to sequences SEQ ID n°8 to 35 described in example 2 below, flanked by the consensus sequences SEQ ID n°6 and 7.

A further subject of the present invention is a rpob gene or gene fragment of a bacterium of genus Streptococcus or of one of said 4 related genera, except sequences SEQ ID n°11, of the bacteria Streptococcus 14, and pyogenes, Streptococcus pneumoniae, Streptococcus mutans and agalactiae, Streptococcus the reverse sequences and complementary sequences, characterized in that it comprises a sequence such as described in sequences SEQ ID  $n^{\circ}$  8 to 35 described in example 2.

A further subject of the invention is the complete sequence of the *rpoB* gene of the bacteria *Streptococcus anginosus*, *Streptococcus equinus* and *Abiotrophia defectiva* such as described in sequences SEQ ID n°1 to 3, which can be used in particular for a method of the invention.

A further subject of the present invention is the almost complete sequence of the *rpob* gene of the bacterium *Enterococcus faecalis* such as described in SEQ ID n°5, which can be used in particular for a method of the invention.

In sequences SEQ ID  $n^{\circ}$  1 to 3 and 5 and 8 to 35 described in the sequence listing at the end of the description:

- nucleotide M represents A or C,
- nucleotide K represents T or G,
- or G,

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- nucleotide W represents A or T,
- nucleotide Y represents C or T,
- nucleotide N represents A, T, C, G or I

- nucleotide S represents C or G,

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- nucleotide V represents A,C or G

The consensus sequences derived from SEQ ID  $n^{\circ}$  6 and 7 evidenced according to the present invention, may be used as genus probe, as amplification or sequencing reaction primer in detection methods for bacteria of genus Streptococcus and said 4 related genera, by molecular identification.

With the sequences derived from sequences SEQ. ID n° 6 7 it is therefore not only possible to prepare genus probes for bacteria of genus Streptococcus and said 4 related genera, but also to detect and identify the species of said bacteria through amplification and sequencing using sequences as primers.

The complete sequence of the rpoB gene may be used to identify the bacterium not only through the study of its primary sequence, but also through the study of the secondary and tertiary structures of the messenger RNA derived from transcription of the complete DNA sequence.

A further subject of the invention is an oligonucleotide or an rpoB gene fragment having a sequence included in or consisting of sequences SEQ ID n° 8 to 35, hence including ID n° 11, sequences SEQ 12, 14 and 22 of the bacteria Streptococcus pyogenes, Streptococcus pneumoniae, Streptococcus and mutans Streptococcus agalactiae 25 respectively, and also among the oligonucleotides or fragments of reverse or complementary sequences such as defined above.

The inventors, after analysing the different sequences and comparing pair by pair all sequences SEQ. ID n° 8 to 35, determined that the homology rate between two different sequences among said sequences SEQ ID n° 8 to 35 is a maximum rate of 98.7% Below 98.7% homology between the sequences, they identify bacteria of different species. Consequently, further subject of the invention is oligonucleotides or rpoB gene fragments having sequences included in or consisting of said sequences SEQ ID n° 8 to 35, the reverse sequences, the complementary sequences and sequences showing at least 98.7% homology (i.e. a similarity rate of at least 98.7% between the sequences) with respect to said sequences SEQ ID n° 8 to 35, the reverse sequences and complementary sequences respectively.

The oligonucleotides, gene fragments and genes subject of the present invention have been described as comprising DNA sequences i.e. with nucleotides A, T, C and G. However, a further subject of the present invention is oligonucleotides comprising corresponding RNA sequences, i.e. in which T is replaced by U.

In the present description, by "reverse sequences and complementary sequences" is meant the following sequences:

- the reverse sequence of said sequence,

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- the complementary sequence of said sequence, and
- the complementary sequence of the reverse sequence of said sequence.
- Sequences SEQ ID  $n^{\circ}$  1 to 35 may be prepared by genetic engineering and/or chemical synthesis, in particular by automatic synthesis, using techniques well known to persons skilled in the art.

One first application of an oligonucleotide of the invention is its use a probe for the detection, in a biological specimen, of bacteria of one of the species of genus *Streptococcus* and said 4 related genera, which comprises a nucleotide sequence in one of the sequences SEQ ID n°6 to 35 and their reverse or complementary sequences.

An oligonucleotide comprising sequences SEQ ID n° 6 and 7 will be used as genus probe, and an oligonucleotide comprising a sequence included in or comprising one of sequences SEQ ID n° 8 to 35 will be used as species probe.

More particularly, the subject of the present invention is an oligonucleotide comprising a sequence specific to a bacterium species of genus *Streptococcus* and said related genera, preferably having at least 20 consecutive nucleotides, further preferably at least 30 consecutive nucleotides included in one of said sequences SEQ ID n° 8 to 35, or optionally an equimolar mixture of said oligonucleotides having different sequences.

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Preferably, said sequences included in one of sequences SEQ ID n° 8 to 35, preferably having at least 20, further preferably at least 30 consecutive nucleotides included in one of said sequences SEQ ID n° 8 to 35, form the shortest sequences specific to the different respective species which can be used as species probe for *Streptococcus* bacteria and for said 4 related genera under consideration.

The probes of the invention may be used for diagnostic purposes, mentioned previously, by determining as formation or non-formation of a hybridisation complex between the probe and a target nucleic acid in the specimen, using all "DOT-BLOT" known hybridisation techniques in particular techniques [Maniatis et al. (1982) Molecular Cloning, Cold Spring Harbor] DNA transfer techniques called "SOUTHERN BLOT" [Southern E.M., J. Mol. Biol. (1975) 98: 503], RNA transfer techniques called "NORTHERN BLOT", or so-called "sandwich" techniques in particular using a capture probe and/or a detection probe, said probes being able to hybridise with two different regions of the target nucleic acid, and at least one of said probes (generally the detection probe) being able to specific to the hybridise with a target region that is species, the capture probe and the detection probe evidently having nucleotide sequences that are at least different.

The nucleic acid to be detected (target) may be DNA or RNA (the first obtained after PCR amplification). When detecting a target of double strand nucleic acid type, the latter must first be denatured before starting detection. The target nucleic acid may be obtained using known methods for its extraction from a specimen to be examined. Denaturing of a double strand nucleic acid may be conducted using known chemical, physical or enzymatic methods, in particular by heating to an appropriate temperature, of over 80°C.

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To implement the above-mentioned hybridisation techniques, in particular the "sandwich" techniques, a probe of the invention called a capture probe is immobilised on a solid support, and another probe of the invention called a detection probe is labelled with a marking agent. Examples of supports and marking agents are those previously given.

Advantageously, a species probe is immobilised on a solid support, and another species probe is labelled with a marking agent.

of Another application an oligonucleotide of the 20 its use as nucleotide primer invention is comprising a monocatenary oligonucleotide chosen from oligonucleotides having a sequence of at least 12 nucleotide motifs included in one of sequences SEQ ID n° 6 to 35, which can be used in the synthesis of a nucleic acid in the presence 25 of a polymerase using a known method, in particular by amplification methods using said synthesis in the presence of a polymerase (PCR, RT-PCR, etc). In particular, a primer of invention may be used for the specific reverse transcription of a messenger RNA sequence of a bacterial 30 species of genus Streptococcus and said 4 related genera to corresponding complementary DNA obtain sequence. reverse transcription may form the first stage of the RT-PCR technique, the following stage being PCR amplification of the

complementary DNA obtained. Primers of the invention may also be used for specific amplification, by chain polymerisation reaction, of the total DNA sequence of the *rpoB* gene of a species of genus *Streptococcus* and said 4 related genera.

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In one particular case, said primer comprising an oligonucleotide of the invention also comprises the sense or antisense sequence of a promoter recognized by a polymerase RNA (promoters T7, T3, SP6 for example [Studier FW, BA Moffatt (1986) J. Mol. Biol. 189:113]: said primers can be used in nucleic acid amplification methods using a transcription step such as, for example, NASBA or 3SR techniques [Van Gemen B et al. Abstract MA 1091, 7th International Conference on AIDS (1991) Florence, Italy].

A further subject of the invention is a nucleotide primer comprising an oligonucleotide chosen from among oligonucleotides having a sequence comprising one of sequences SEQ ID  $n^{\circ}$  6 to 35 or a sequence included in SEQ ID  $n^{\circ}$  6 to 35 which can be used for full or partial sequencing of the rpoB of any strain of а bacterial species of Streptococcus and said 4 related genera.

Full or partial sequencing of the *rpoB* gene in any bacterium of genus *Streptococcus* and related genera enables the identification of all bacteria of genus *Streptococcus* and of said 4 related genera by bio-computerized analysis of this sequence, and enables the recognition of new unknown bacterial species of *Streptococcus* and of said 4 related bacteria.

Preferably, for use as a primer or for sequencing rpoB genes, said mixture of oligonucleotides of the invention is used, and further preferably said mixtures of oligonucleotides consisting of sequences SEQ ID  $n^{\circ}$  6 and SEQ ID  $n^{\circ}$  7.

More precisely, the present invention provides a detection method by identification to detect a bacterium of

one of the species of genus *Streptococcus* and of said 4 related genera, characterized in that the following are used:

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- a complete or almost complete rpoB gene of said bacterium according to the present invention and an rpoB gene or gene fragment of a bacterium Streptococcus pyogenes, Streptococcus Streptococcus pneumoniae, mutans and Streptococcus agalactiae comprising a sequence such as in sequences SEQ ID n° 11, 12, described 14 reverse sequences and complementary respectively, the sequences, which may be used in particular as species probe, and/or
- a said fragment of said rpoB gene of said bacterium to the present invention, comprising nucleotide sequence chosen from among one of sequences SEO n° to 35, the reverse sequences complementary sequences, which may be used in particular as species probe, and/or
- an oligonucleotide of the present invention comprising a sequence included in one of sequences SEQ ID n°8 to 35, the reverse sequences and complementary sequences, which may be used in particular as species probe, and/or
- an oligonucloetide or said mixture of oligonucleotides of the present invention comprising a sequence consisting of consecutive nucleotide motifs, included in one of sequences SEQ ID  $n^{\circ}$  6 and 7, which may be used in particular as genus probe or amplification primer.

Preferably, in said detection method of the invention, the following are used:

a said rpoB gene fragment of said bacterium comprising a
 sequence chosen from among one of sequences SEQ ID n° 8
 to 35 or an oligonucleotide having a sequence included in one of said sequences SEQ ID n° 8 to 35, the reverse sequences and complementary sequences, and/or

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- at least one said mixture of oligonucleotides according to the present invention whose preferable sequences consist of sequences SEQ ID n° 6 and 7, and their reverse sequences and complementary sequences in which further preferably N represents inosine.

In a first embodiment of a detection method of the invention, it is sought to evidence the presence of a bacterium of genus *Streptococcus* and said 4 related genera, and the following steps are performed in which:

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- 1. at least one genus probe comprising a said mixture of oligonucleotides having sequences comprising or included in one of sequences SEQ ID n° 6 and 7, the reverse sequences and complementary sequences according to the invention, is contacted with a specimen containing or possibly containing nucleic acids of at least one said bacterium of genus Streptococcus and of said 4 related genera, and
  - 2. the formation or non-formation is determined of a hybridisation complex between said genus probe and the nucleic acids of the specimen, and the presence is determined of said bacterium of genus *Streptococcus* or of said 4 related genera if a hybridisation complex is formed.

In a second embodiment of a detection method for a 25 bacterium of genus *Streptococcus* and said 4 related genera, the steps are performed in which:

1. Amplification primers, comprising said mixtures of oligonucleotides containing a sequence included in said SEQ ID n° 6 and 7 reverse sequences sequences complementary sequences of the invention, are contacted with a sample containing or possibly containing nucleic acids of at least one said bacterium of genus Streptococcus and of said 4 related genera, using:

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- as 5' primer: a said mixture of oligonucleotides containing a sequence included in sequence SEQ ID n° 6 or preferably consisting of said complete sequence SEQ ID n°6, or a complementary sequence of the invention,
- as 3' primer: a said mixture of oligonucleotides containing a sequence included in sequence SEQ ID n° 7 or preferably consisting of said complete sequence SEQ ID n°7, or respectively a complementary sequence of the invention.
- 2. The nucleic acids are amplified by enzymatic polymerisation reaction, and the occurrence or non-occurrence of an amplification product is determined, and in this way the presence is determined of said bacterium in the specimen if an amplification product is produced.

This second embodiment may be used to specifically detect the genus of a Streptococcus bacterium or said 4 related genera.

However, at step 2 of this second embodiment, it may be 20 sought to specifically detect a given bacterium species of from Streptococcus chosen among the species genus Streptococcus mutans, Streptococcus oralis, Streptococcus pneumoniae, Streptococcus pyogenes, Streptococcus salivarius, sanguinis, Streptococcus suis, Streptococcus Streptococcus agalactiae, 25 acidominimus, Streptococcus Streptococcus anginosus, Streptococcus constellatus, Streptococcus difficilis, Streptococcus dysgalactiae, Streptococcus equi, Streptococcus equinus, Streptococcus intermedius, Streptococcus mitis, Streptococcus bovis, Granulicatella 30 adjacens, Abiotrophia defectiva, Enterococcus avium, casselliflavus, Enterococcus Enterococcus faecalis, Enterococcus faecium, Enterococcus gallinarum, Enterococcus sacharolyticus, Gemella haemolysins and Gemella morbillorum,

as described in the variant of embodiment of a detection method specific to a species of said bacteria, given in the description below.

As previously set forth in the introduction, the genera Streptococcus, Enterococcus, Granulicatella, Abiotrophia and Gemella comprise more bacterial species than those effectively sequenced in this work. However, the sequenced species were chosen so that they encompass all known species in these bacterial genera and are sufficient in number to demonstrate the application of the rpoB sequence to the identification of the species of these genera.

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In a variant of embodiment of a method of the invention for specifically detecting a species of said bacteria, the steps are performed in which:

- 1. a specimen containing or possibly containing nucleic acids of at least one said bacterium is contacted with at least one species probe consisting of said gene, said gene fragment or said oligonucleotide containing a sequence included in one of sequences SEQ ID n° 8 to 35, preferably an oligonucleotide consisting of one of said sequences SEQ ID n° 8 to 35, the reverse sequences and complementary sequences according to the invention, and
  - 2. the formation or non-formation of a hybridisation complex is determined between said probe and the nucleic acids in the specimen, thereby determining the presence of said bacterium in the specimen if a hybridisation complex is formed.

In another variant of embodiment of the method of the invention, in which it is sought to specifically detect a given species of a bacterium of genus *Streptococcus* and of said 4 related genera, chosen from among the 28 species cited above, the method comprises the steps in which, in a specimen

containing or possibly containing nucleic acids of at least one said bacterium:

a) a sequencing reaction is conducted of an amplified *rpoB* gene fragment of said given bacterium using nucleotide primers consisting of said mixtures of oligonucleotides containing sequences included in sequence SEQ ID n° 6 as 5' primer, and in SEQ ID n° 7 as 3' primer, the sequences preferably consisting of said sequences SEQ ID n° 6 and 7, and their complementary sequences, and

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- 10 b) the presence or absence of the given species of said bacterium determined by comparing is the obtained sequence of said fragment with the sequence of complete rpoB gene of said bacterium or the sequence of a rpoB gene fragment of said bacterium containing said 15 sequences n°8 to 35 and complementary sequences of the invention, thereby determining the presence of said bacterium in the specimen if the obtained fragment sequence is identical to the known sequence of the genus or of the rpoB gene fragment of said bacterium.
- 20 A further subject of the present invention is a diagnosis kit which can be used for а method of the invention, containing at least one said gene fragment or oligonucleotide having a sequence included in or consisting of sequences SEQ ID n° 8 to 35, or a said oligonucleotide or 25 mixture of oligonucleotides containing a sequence included in one of sequences SEQ ID n° 6 and 7, and/or at least one said rpoB gene fragment of said bacterium comprising sequences SEQ ID n° 8 to 35 and complementary sequences of the invention.

Advantageously, a kit of the present invention contains 30 said oligonucleotides in the form of "biochips", i.e. fixed to solid supports, glass in particular, according to the method described in US patent 5,744,305 (Affymetrix, Fodor et al) using the resequencing strategy described in application WO

95/11995 (Affymax, Chee et al) or according to the method described by A. Troesch et al. in J. Clin. Microbiol., vol. 37(1), p 49-55, 1999. The oligonucleotides synthesized on the "biochip" carry out re-sequencing of the hyper variable region of the rpoB gene. This method offers considerable advantage in terms of production costs with no detriment to quality of identification of the different species through the choice of these identification sequences. Preferably, oligonucleotides fixed onto the "biochip" solid comprise 10 to 30 bases, e.g. 20 bases, with an interrogation position located in the central region for example at position respect to the 3*'* end of the sequence oligonucleotides with 20 bases. Another example consists of using oligonucleotides having 17 bases with 2 interrogation positions: one at position 10 and one at position 8. Other oligonucleotides have lengths of 10 25 between and nucleotides. The interrogation positions then vary according to the length of the oligonucleotide.

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Analysis is conducted on the complete GeneChip® system 20 (reference 900228, Affymetrix, Santa Clara, CA) which comprises the GeneArray® reader, the GeneChip® hybridisation GeneChip® fluid station oven, and GeneChip® analysis software.

An oligonucleotide of the invention may also be used as a gene therapy probe to treat infections caused by a strain belonging to a species of genus *Streptococcus* and said 4 related genera, said probe comprising an oligonucleotide such as defined previously. This gene therapy probe, able to hybridise on the messenger RNA and/or on the genomic DNA of said bacteria, may block translation and/or transcription and/or replication phenomena.

The principle of gene therapy methods is known and is based in particular on the use of a probe corresponding to an

antisense strand: the formation of a hybrid between the probe and the sense strand is able to disrupt at least one of the genetic information decoding steps. Gene therapy probes can therefore be used as anti-bacterial medicines, making it possible to fight against infections caused by bacteria belonging to the species of genus *Streptococcus* and said 4 related genera.

The invention will be more readily understood with the help of the description given below, divided into examples relating to experiments conducted with a view to implementing the invention and which are given solely for illustrative purposes.

Figure 1 shows the visualisation of the amplification products through ethidium bromide staining after electrophoresis on an agarose gel obtained in example 3.

Example 1: Sequence of the *rpoB* gene of three species of genus Streptococcus and related genera: Abiotrophia defectiva, Streptococcus anginosus and Streptococcus equinus.

20 The complete sequence of the rpoB gene of bacteria belonging to the species of Abiotrophia defectiva, Streptococcus anginosus and Streptococcus equinus determined by enzymatic amplification and automatic sequencing available for Streptococci. The choice of these species was 25 on analysis of the 16S tree which based shows divergence covering the entire phylogenetic tree streptococci.

Strategy and Sequencing:

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Several partial 510-bp sequences of rpoB genes are available from GenBank for the 10 following streptococcus species: Streptococcus intermedius, Streptococcus sanguinis, Streptococcus penumoniae, Streptococcus parasanguinis, Streptococcus oralis, Streptococcus mitis, Streptococcus

cristalus, Streptococcus constellatus, Streptococcus anginosus, and Granulicatell adjacens [Majewski J., Zawadzki Pickerill P., Cohan F.M. and Dowson C.G. Barriers to P., genetic exchange between bacterial species: Streptococcus pneumoniae transformation. J. Bacteriol. 182, 1016-1023 (2000)], but the primers used by these authors only amplify a fraction of the species of genus Streptococcus, and it was therefore not possible to carry out our work on the basis of this data alone. It was therefore necessary to determine primers able to amplify all strains of streptococci, enterococci, Abiotrophia, Gemella and Granulicatella. These primers also had to flank a region showing sufficient genetic diversity so as to be able to distinguish between two species. However, the alignment of these published partial sequences 15 made it possible to determine the following common primers: refers (the numbering to the complete sequence of Streptococcus pyogenes)

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SEQ ID n° 36: 5'- AGACGGACCTTCTATGGAAAA-3' (primer 748F) SEQ ID n° 37: 5'- GGACACATACGACCATAGTG-3' (primer 116R), and SEQ ID n° 38: 5'- GTTGTAACCTTCCCAWGTCAT -3' (primer 830R).

These primers allowed the sequencing of the central part of the rpoB gene with 714 bp for the five chosen species (Streptococcus equinus, Streptococcus mutans, Streptococcus anginosus, Enterococcus faecalis, and Abiotrophia defectiva. From this central fragment, sequencing was continued using the so-called genome Walker technique.

Outside this published zone [Majewski J. et al, J. Bacteriol. 2002, 182, 1016-1023], the alignment of the two complete sequences available from GenBank (Streptococcus pneumoniae [GenBank access number AE008542] and Streptococcus pyogenes [GenBank access number AE006480] made it possible to choose the following primers:

-SEQ ID n° 39: 5'- GTCTTCWTGGGYGATTTCCC-3' (primer 2215R)

- -SEQ ID n° 40:5'- ACCGTGGIGCWTGGTTRGAAT-3' (primer 2057R)
- -SEQ ID n° 41: 5' AACCAATTCCGYATYGGTYT-3' (primer 1252R)
- -SEQ ID n° 42: 5' AGIGGGTTTAACATGATGTC-3' (primer 371F)
- -SEQ ID n° 43: 5' AGIGCCCAAACCTCCATCTC-3' (primer 730F), and
- 5 SEQ ID n° 44: 5' CTCCAAGTGAACAGATGTGTA-3' (primer 585R)

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With these primers, it was possible to extend the sequenced region for some of the five chosen strains. In fully unexpected manner, *E. Faecalis* is not amplified by these primers; but it was observed that the sequenced partial zone showed homology with the *rpoB* gene of *Listeria monocytogenes*, i.e. with a bacterium belonging to a different bacterial genus which could in no way be inferred from existing data, and we therefore chose primers in the *rpoB* gene of *Listeria* to amplify the *rpoB* gene of *Enterococcus faecalis*.

- -SEQ ID n° 45: 5'-TTACCAAACTTAATTGAGATTCAAAC-3' (primer 180F)
  - -SEQ ID n° 46: 5' AGTATTTATGGGTGATTTCCCA-3' (primer 410F)
  - -SEQ ID n° 47: 5'- GGACGTTATAAAATCAACAAAAATT-3' (primer 910F)
  - -SEQ ID n° 48:5'- AGTTATAACCATCCCAAGTCATG-3' (primer 2430R)
  - -SEQ ID n° 49:5'- TGAAGTTTATCATCAACCATGTG-3' (primer 3280R)
- 20 SEQ ID n° 50: 5' CCCAAAACGTTGTCCACC-3' (primer 3360R)

The partial sequences so obtained for the five chosen strains (Streptococus equinus, Streptococcus mutans, Streptococcus anginosus, Enterococcus faecalis, Abiotrophia defectiva) made it possible to choose the following primers:

- 25 -SEQ ID n°51: 5'- AACCAAGCYCGGTTAGGRAT-3' (primer 520R)

  -SEQ ID n°52: 5'- ATGTTGAACCCACTIGGGGTGCCAT-3' (primer 2881F)

  for the sequencing of the end C- and N- zones by Genome Walker.
- Sequencing was then complete as displayed by the determination of the encoding region and the alignment of the translated proteins of the nucleotide sequences with the two published rpoB proteins of Streptococcus pneumoniae and Streptococcus pyogenes.

Several potential consensus primers were investigated to obtain a fragment able to lead to the complete sequence of the rpoB genes by successive elongations from a series of specific primers.

In each of the above steps, a large number of attempts with theoretically or potentially suitable primers failed before the above-mentioned primers were determined enabling the amplification and sequencing in successive steps of the entirety of the *rpoB* genes described below.

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The sequencing reactions were conducted using reagents from the kit: ABI Prism dRhodamine Dye Terminator Sequencing Ready Reaction Kit (Perkin Elmer Applied with Biosystems) in accordance the manufacturer's recommendations and following the programme: 30 comprising a denaturing step at 94°C for 10 sec., hybridisation step of the primer at 50°C for 10 sec. and an extension step at 60°C for 2 minutes. The sequencing products were separated by electrophoresis on a polyacrylamide gel and 377 DNA sequencer (Perkin) and analysed to form consensus sequences using Sequence Assembler software (Applied Biosystems).

With this approach we were able to determine the complete sequence of the *rpoB* gene in two species of genus Streptococcus and in Abiotrophia defectiva:

SEQ ID n° 1: Sequence of the *rpoB* gene of *Streptococcus* anginosus. This sequence measures 4523 base pairs, has a guanosine plus cytosine content of 41% and is deposited in GenBank under accession number AF 535183:

5'-tcatacttttagagtcagatttagctgctctttttgtgcctgttttgggatttttgtcgtttgt CATCAAAATTAAAGATTCTGAAAATTACTCAAAAAGGATAAATGAAAATTGCTACTCTATTCCA TTAATAGAGAATGTAGAAAGAAGAAGGAGTAAAAAACTTGGCAGGACATGAAGTTCAATACGGG AAACACCGTACTCGTCGTAGTTTTTCAAGAATCAAGGAAGTTCTTGATTTACCAAATTTGATTG AAATCCAGAGGATTCGTTCAAAGATTTTCTTGACCATGGTTTGAAAGAAGTATTTGAAGATGTA CTTCCTATCTCAAACTTTACAGATACAATGGAGCTAGAGTTTGTTGGTTATGAAATTAAAGGAT CTAAATACACTTTAGAAGAAGCACGTATCCATGATGCCAGCTATTCTGCACCTATTTTTGTGAC TTTCCGTTTGATTAATAAGAAACTGGTGAAATCAAAACCCAAGAAGTGTTCTTTGGCGATTTC CCAATCATGACAGAAATGGGAACTTTCATTATCAATGGTGGTGAGCGGATTATCGTATCTCAGC TCGTTCGTTCTCCAGGTGTTTACTTCAACGATAAAGTAGACAAAAATGGTAAAGTTGGTTATGG TTCAACTGTCATTCCTAACCGTGGAGCTTGGTTAGAGCTGGAAACAGACTCAAAAGATATTGCT TATACTCGGATTGACCGTACTCGTAAGATTCCGTTTACGACACTTGTTCGTGCGCTTTGTTTT CTGGCGATGATGAAATCTTTGACATTTTCGGCGACAGCGATCTCGTTCGCAACACGATTGAAAA GGATATTCATAAAAATCCAATGGATTCACGTACGGATGAAGCGCTTAAAGAAATCTATGAACGT CTTCGTCCAGGTGAGCCTAAAACAGCTGATAGTTCACGTAGTCTATTGGTCGCTCGTTTCTTTG ATCCACATCGTTACGACTTGGCGGCAGTTGGTCGTTATAAAAATCAATAAAAAATTAAACATTAA AACACGTTTGTTAAATCAAACGATTGCAGAGCCTTTGGTAGATCCAGAAACAGGTGAAATCTTG GTTGAAGCTGGAACGGTTATGACGCGTAGTGTCATTGATAGCATTGCAGAATACTTGGACGGTG ATTTGAATAAAATCACTTATATTCCAAATGATGCAGCTGTGTTAACAGAGCCAGTTGTTCTTCA AAAATTCAAAGTGGTGGCGCCAACTGATCCAGATCGTGTGGTGACTATTATTGGTAATGCCAAC CCAGGAGATCGAGTTCATACGATTACGCCAGCAGATATTTTGGCTGAGATGAATTACTTCTTGA ACCTCGCTGAAGGACTTGGTCGTGTGGACGATATTGACCACTTGGGAAATCGTCGGATTCGTGC CGTTGGTGAATTGCTTGCTAACCAAGTACGTCTTGGCTTGTCTCGTATGGAGCGAAACGTTCGG GAGCGCATGAGTGTGCAAGATAATGAAGTGTTGACACCGCAACAATCATTAACATCCGCCCAG TCACAGCAGCTATCAAAGAATTCTTTGGTTCATCTCAATTGTCTCAATTTATGGACCAACATAA TCCACTGTCTGAATTGTCTCACAAACGCCGTTTGTCAGCCTTGGGACCTGGTGGTTTGACTCGT  ${\tt GATCGTGCTGGATATGAAGTGCGTGACGTGCACTATACCCACTATGGTCGTATGTGTCCGATTG}$ AAACGCCTGAAGGACCAAACATCGGTTTGATCAATAACTTGTCTTCTTATGGACACTTGAATAA ATATGGCTTTATCCAAACGCCGTATCGTAAAGTGGATCGTGAAACAGGTCTGGTCACCAATGAA CAGAAGATGGTCGTTTTGCAGAAGCGATTGTCATGGGACGTCACCAAGGGAACAACCAAGAATT TCCTTCAGATCAAGTAGACTTCATGGATGTATCGCCTAAGCAGGTAGTTGCGGTTGCGACAGCA TGTATTCCTTGAAAACGACGACTCAAACCGTGCTCTCATGGGTGCCAACATGCAACGTC AGGCGGTACCGTTGATTGATCCGCATGCACCATATGTTGGTACTGGTATGGAATACCAAGCAGC TCATGACTCTGGTGCGGCGATTATTGCCCAACACGACGGTAAAGTTGTATATTCTGATGCAGCC AAAGTTGAAGTTCGTCGTGAAGATGGCTCACTTGATGTCTATCATATTACGAAATTCCGCCGTT CAAACTCTGGTACTTCTTACAACCAACGTACGCTGGTAAAAGTTGGCGATACAGTTGAAAAAGG TGACTTTATCGCAGACGGACCTTCTATGGAAAAAGGTGAAATGGCACTTGGACAAAATCCAATC TGAAAGACGATGTTTACACATCTGTTCACTTGGAGGAATTTGAATCAGAAACACGTGATACAAA STRF GCTTGGACCTGAAGAATCACGCGCGAAATTCCAAACGTCGGTGAAGATGCTTTGAGAGACCTT GACGAAACGGGAATTATCCGCATTGGTGCTGAGGTAAAAGAAGGCGACATTCTTGTCGGTAAAG TAACACCGAAAGGTGAAAAAGACTTATCTGCTGAAGAACGCCTGCTTCATGCAATTTTTCGGTGA TAAATCTCGTGAAGTACGTGATACTTCCCTTCGTGTACCACATGGTGGTGCAGGGGTTGTCCGT GATGTGAAAATCTTTACTCGTGCGAACGGTGATGAATTGCAATCTGGTGTCAACATGTTGGTAC GTGTTTACATCGCTCAAAAACGGAAAATCCGTGTTGGGGATAAGATGGCTGGACGTCACGGAAA

CAAAGGGGTTGTTTCCCGCATTGTTCCAGTTGAGGATATGCCGTATCTTCCAGATGGAACACCA

GTTGATATTATGTTGAACCCACTTGGGGTGCCATCTCGTATGAATATTGGTCAAGTTATGGAGC TTCACCTCGGTATGGCTGCTCGCAACCTTGGCATTCACATTGCAACACCAGTATTTGACGGGGC TAGCTCAGATGATCTTTGGGAAACCGTTCGTGAAGCTGGCATGGATAGCGATGCTAAGACAATC CTTTATGATGGCCGTACTGGTGAGCCATTTGATAATCGTGTATCCGTTGGTGTCATGTACATGA TCAAACTCCACCATATGGTTGATGATAAGCTCCATGCCCGTTCCGTTGGTCCTTATTCAACCGT STRR TACGCAACAACCTCTTGGTGGTAAAGCGCAGTTTGGTGGACAACGTTTTGGAGAAATGGAAGTT TGGGCTCTTGAAGCCTACGGTGCTTCTAACGTCCTTCAAGAAATCTTGACTTACAAGTCAGATG ACATCAATGGTCGTTTGAGAGCTTATGAAGCCATTACCAAAGGTAAGCCAATTCCAAAACCAGG TGTTCCAGAATCCTTCCGTGTCCTTGTAAAAGAATTGCAATCACTTGGTCTTGACATGCGTGTC CTTGATGAAGACGACAATGAAGTCGAACTTCGTGACTTGGACGAAGGCATGGATGATGATGTGA TTCATGTAGACGATCTTGAAAAAGCACGTGAAAAAGCAGCACAAGAAGCAAAAGCCGCTTTTGA ATGTAAATCGTTTTCAAAGTATGCAAATCACCCTAGCTTCTCCTAGTAAAGTCCGCTCTTGGTC TTATGGAGAAGTGAAGAAACCTGAAACAATTAACTACCGCACACTAAAACCAGAACGCGAAGGG CTTTTTGATGAAGTCATCTTTGGTCCTACGAAAGACTGGGAATGTGCGTGTGGAAAATATAAAC GGATTCGTTATAAAGGAATCATTTGTGACCGTTGTGGTGTTGAAGTAACTCGTACTAAAGTTCG TCGTGAACGTATGGGACATATTGAGTTGAAAGCCCCAGTCTCCTCATATTTGGTATTTTAAAGG AATTCCAANTCGCATGGGCTTGACCTTGGACATGAGCCCTCGTGCTCTTGAAGAAGTCATNTAN TTTGCAGCTTATGTGGTGANTGACCCTAAAGATACNCCACTTGAGCACAAATCCATTATGACAG AGCGGGATGGTTNGTGAACGCTGACNTGAATATGGCCAAGGCTCTTTTGTTGCAAAAATGGGTG YTGAAGCAATCCAAGATCTNNTGAAACANGTAGACCTGGAAAAAGAAATTGCAGAGCTCAAAGA TGAATTAAAAACGGCAAGTGGGCAAAAGCGCGTAAAMGCTAANTTCGTCGNTNNGACTCTTTTC GATNCTTTCCAAAAATCATGGTACACAAAACCAGAACTGGATGGTCTTAAACCATCNTNTCACC GCTCATTCCAGACAC -3'

SEQ ID n° 2: Sequence of the *rpoB* gene of *Streptococcus* equinus. This sequence measures 4118 base pairs, has a guanosine plus cytosine content of 41% and is deposited in GenBank under number GenBank accession AF 535187:

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5'-CACGCGTGGTCGACGGCCCGGGCTGGTGAATTGTCATAAGTTGTGTAGTAGTAAATTCCCTTAT CAGTGTTGATGCATGAGCTATAAATAGTGTACTCATATTTGCCACTTTCATCGACATAGCAAAG TCCTTTTTGTTGTTCAACGGATTTTAAAATGTGGAAGAATTGATTAACACTGCTTTCTTCTTGTT TCTTCAGCCACAGAATTTAATTTTGTAAAAGTAACTTTTACATAACGTGACATTGATGATAAAT CACCAGGCAAGCCAAGTCCACCCATGCCACGGCTATAAGTTTCAAGTTCTAACTCTTTAGCAAA ACGATTTTCTGAAACCTTTGGAGATAGATGACGATAGTTATTCAAATTGAATAATTGTTTATCA AAAGTTGGATTATTAGTCAAAACACCTGTTGAGTTATTCGTAAACTTATAGGGCACGCGTGGTC GACGGCCCGGGCTGGTAAAGACTTCTTGGATAACGGATTAAMAGAAGTTTTTGAAGATGTACTT CCGATTACAAACTTTACGGATACTATGGAGCTTGAATTTGTTGGTTACGAATTGAAAGAGCCTA AGTATACGCTTGAAGAAGCTCGTATCCACGATGCATCTTATTCAGCACCTATTTTTGTAACCTT CCGTTTGATTAATAAAGAAACAGGAGAAATCAAAACTCAAGAAGTTTTCTTCGGTGATTTCCCA ATTATGACTGAAATGGGTACATTCATCATCAACGGTGGTGAACGTATTATCGTTTCTCAGTTGG TTCGTTCTCCTGGTGTTTATTTCAACGATAAAGTTGATAAAAACGGTAAAGTTGGTTACGGTTC AACTGTAATCCCTAACCGTGGAGCATGGCTTGAATTAGAAACAGATTCAAAAGATATTGCTTAC ACACGTATCGACCGTACACGTAAAATTCCATTTACAACTCTTGTACGTGCGCTTGGTTTCTCAG GTGATGAAAATCATGGATATCTTTGGTGATAGCGAACTTGTTCGTAACACAATCGAAAAAGA TATTCACAAAAACCCAGCAGACTCACGTACTGACGAAGCTCTTAAAGAAATTTACGAACGCCTT CACGTCGTTATGACTTGGCAGCTGTTGGTCGTTACAAAATCAACAAAAAACTTAACATCAAGAC TCGTCTTTTGAACCAAACAATCGCTGAAAACTTGGTTGATGCTGAAACTGGTGAAATCCTTGTT GAAGCTGGTACAGTAATGACACGTGACGTGATTGATTCAATCGCTGATCAATTGGATGGTGACC TTAACAAATTTGTTTACACACCAAATGATTACGCTGTTGTCACTGAACCTGTTGTTCTTCAAAA ATTCAAAGTTGTTGCACCAAACGATCCAGACCGCGTTGTTACAATCGTTGGTAACGCAAATCCT GATGACAAAGCGCGTGCGCTTACACCAGCTGATATCTTGGCAGAAATGTCTTACTTCCTTAACC TTGCTGAAGGTCTAGGTAAAGTTGATGATATCGACCACCTTGGGAATCGTCGTATTCGTGCCGT TGGTGAATTGCTTGCTAACCAATTCCGTATTGGTCTTGCTCGTATGGAACGTAACGTTCGGGAA CGTATGTCAGTTCAAGACAACGAAGTGTTGACACCACAACAATCATCAACATTCGTCCTGTTA

CTGCAGCCGTTAAAGAATTCTTCGGTTCATCTCAATTGTCACAGTTCATGGACCAACACACCC ACTTTCTGAGTTGTCTCACAAACGTCGTTTGTCAGCCTTAGGACCTGGTGGTTTGACTCGTGAC TGGTTTCATCCAAACACCATATCGTAAAGTTGACCGCGCTACAGGTGTGATTACAAACGAAATC GTTTGGTTGACTGCCGATGAAGAAGATGAATACACAGTAGCACAGGCTAACTCAAAACTTAACG AAGATGGAACATTTGCTGAAGACATCGTTATGGGACGTCACCAAGGTAATAACCAAGAGTTCCC AGCAAGCGTTGTTGACTTCGTAGACGTTTCACCTAAACAAGTAGTTGCCGTTGCGACAGCATGT ATTCCTTTCCTTGAAAACGATGACTCTAACCGTGCCCTTATGGGTGCCAACATGCAACGTCAAG CGGTGCCATTGATTGATCCACACGCACCATATGTTGGTACTGGTATGGAATATCAAGCAGCCCA CGACTCAGGTGCTGCAGTTATCGCTAAACACGATGGACGCGTTATCTTCTCTGATGCTGAAAAA GTTGAAGTTCGTCGCGAAGATGGTTCACTTGATGTTTACCACATTACTAAATTCCGTCGTTCTA ACTCAGGTACAGCTTATAACCAACATACACTTGTTAAAGTTGGCGATATCGTTGAAAAAGGTGA CTTCATCGCTGATGGTCCTTCAATGGAAAAAGGTGAAATGGCCCTTGGTCAAAACCCAATCGTC GCTTACATGACTTGGGATGGTTATAACTATGAAGATGCCATCATCTTGAGTGAACGTCTTGTTA AAGAAGATGTTTATACATCAGTTCACTTGGAAGAATTTGAATCAGAAACACGTGATACTAAGTT STRF AGGCCCTGAAGAATCACTCGCGAAATTCCAAACGTTGGTGAAGAAGCTCTTAAAGACCTTGAC GAAATGGGTATTATCCGTATCGGTGCTGAAGTTAAAGAAGGTGACATCCTTGTAGGTAAAGTAA CACCTAAAGGTGAAAAAGACCTTTCTGCTGAAGAGCGCCTTCTTCACGCAATCTTCGGTGATAA TTTATATCGCACAAAAACGTAAAATCAAAGTCGGAGATAAAATGGCCGGTCGTCACGGTAACAA AGGGGTTGTTCTCGTGTTGTTCCAGTTGAAGACATGCCTTATCTTCCAGACGGAACTCCAGTC GATATCATGTTGAACCCACTTGGGGTGCCATCTCGTATGAACATCGGACAAGTTATGGAGCTTC ACCTTGGTATGGCTGCTCGTAACCTTGGTATTCACATTGCAACACCAGTCTTTGATGGGGCCAAC TTCTGAAGACCTTTGGGATACAGTTAACGAAGCTGGTATGGCTAGCGACGCTAAGACAGTTCTT TACGATGGACGTACTGGTGAACCATTTGATAACCGTGTGTCAGTTGGTGTCATGTACATGATTA AACTTCACCACATGGTTGATGATAAACTTCACGCACGTTCAGTTGGTCCTTACTCACTTGTTAC STRR GCAACAACCTCTTGGTGGTAAAGCACAATTTGGTGGACAACGTTTCGGTGAAATGGAAGTTTGG GCTTTGGAAGCTTACGGTGCATCAAATGTTCTTCAAGAAATCTTGACTTACAAATCAGATGATG TCAACGGTCGTCTTAAAGCTTATGAAGCCATCACTAAAGGTAAACCAATTCCAAAACCAGGTGT TCCAGAATCATTCCGAGTTCTTGTAAAAGAATTGCAATCACTTGGTCTTGACATGCGCGTGCTT GATGAAGATGACAATGAAGTAGAACTTCGTGATCTTGATGAAGGTGAAGATGACGATGTTATGC ACGTTGATGATCTTGAAAAAAGCTCGTCAAAAAACAAGAAGCAGAAGAAGCGGAAAAAAGCAGAAGT TTCTGCAGAAGAAACAATAATAGGAAAGAACATTCAGACATGAGAGAGGCAAGACCTGCTTC TCTTGGTCAGATTGTTTGATTGAGTCCTATAACGATAAATGATGTCTTACGAATCATGAATTTG TAAGTCATGACAGTTAGAAAGTAGCGCAGCTATTTCAAAGTCATAAGAAGGTATCATGGTGACG TAATCGTTACAGCCGGCGTC -3'

SEQ ID  $n^3$ : Sequence of the *rpoB* gene of *Abiotrophia defectiva*. This sequence measures 4325 base pairs, has a guanosine plus cytosine content of 47%, and is deposited in GenBank under number AF 535173:

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TCTTGGTTGCGCGCTTCTTCGACCCACGTCGCTATGACTTAGCACCTGTAGGTCGTTATAAGAT CAATAAAAAGCTCCACCTCAAGAACCGTTTGGTTGGCTTGACTTTGGCTGAAACCTTTGGTTAAC CCAGAAACAGGCGAAGTGCTCTTTGAAGAAGGAACGGTCTTGGATCAAGAACGTGTTCAAGCCC TGATTCCATACTTAGAGGCTGGCTTGAATAAGGTAACCCTCTATCCTTCTGAAGATAGTGTGGT AGCTCAACCAATTGATTTACAAATCATCAAAGTTTATTCACCTAAGAACGCCGAGCAAGTGATT AACATCATCGGTAACGGGAACATTGAGAAGATTAAGTGCTTGACGCCAGCTGACATTATTGCGT CAATGAACTACTATCTCTATTTAGACCAAGGAATTGGTGTGACAGATGATATCGACCACTTGGC TAACCGTCGTATTCGTTCAGTCGGTGAATTATTGCAAAACCAATTCCGTATCGGGCTATCCCGG ATGGAACGGTAGTGCGTGAACGTATGTCGCTCCAAGATGTTGCGACCATCACACCGCAACAAT TGATTAACATTCGTCCAGTAGTGGCGGCTATTAAGGAATTCTTCGGTTCATCCCAGTTGTCACA ATTCATGGACCAAGTTAACCCACTCGGGGAATTGACCCACAAACGTCGTCTGTCAGCCTTAGGG CCTGGTGGTTTGACGCGGGACCGTGCCGGCTATGAAGTGCGGGACGTTCACTACTCTCACTACG GCCGTATGTGTCCAATCGAGACGCCAGAAGGTCCTAACATCGGGTTGATTAACAGCTTGTCTTC TTATGCCAAGATTAACAAGTATGGTTTTATTGAGACGCCTTACCGTAAAGTGGACAAATCGGTT ACGCCACACCGTGTCACGACCGAAATTGACTACCTAGCAGCGGACGAGGAAGACTTGTACGTAG TAGCCCAAGCCAACTCTAAACTCAACGAAGACGGGACCTTCGCCAATGACCTAGTTATGGCGCG TTTCCGTTCACAAAACATTGAGGTTAACGTTGACCAAGTAGACTACATGGACGTATCGCCAAAA  ${\tt CAGGTTGTCGCTGTCGCACTGCTAGCATTCCGTTCTTGGAAAACGACGACTCCAACCGGGGCT}$ TGATGGGTGCCAACATGCAACGTCAAGCTGTGCCACTTATTAATCCACAATCCCCACTGATTGG GACTGGGATGGAATATAAGGCAGCACACGACTCTGGGGCTGCGCTCTTATGTAAGCGCGCCGGT GAAGTGGTTTATGTCGATGCTAACAAGGTGCGCGTGCGCACTCCAGAAGGTGAAGTTGACGAAT ATTAGGCGACCAAGTTGATGCCTTGGAAATCTTAGCAGATGGTCCATCTATGCAAAATGGGGAG ATGGCCCTCGGTCAAAACCCACTGGTAGCCTTCATGACTTGGGAAGGGTATAACTATGAGGACG CGGTTATCATGTCTGAACGTCTGGTCAAAGACGATGTTTATACCTCTATCCACATTGAAGAATA TGAATCAGAGTCCCGTGAYACYAAGTTAGGCCCTGAAGAAATTACACGCGAAATTCCAAACGTG STRF TCCGAAGATGCCCTCAAGTACTTAGACAAAGACGGGATTATCTGTATCGGGGCGGAAGTAAAAG ACGGCGATATCTTAGTTGGTAAGGTAACACCAAAAGGTGTGACCGAGTTGTCTGCGGAAGAACG CTTGCTCCATGCTATCTTCGGTGAGAAGGCGCGTGAAGTACGTGATACTTCCTTGCGTGTGCCA CACGGCGGGGGGGATTGTCCACGACGTTAAAATCTTTACCCGCGAAGCTGGCGACGAATTGG CACCAGGTGTCAACAAGCTAGTCCGCGTCTACATCGTACAAAAACGTAAAATCAATGAAGGGGA TAAGATGGCCGGTCGTCACGGTAACAAAGGGGTTGTCTCCCTTATCATGCCGGAAGAAGATATG CCATTCTTACCAGATGGTACCCCAGTTGATATCATGTTGAACCCATTAGGGGTTCCATCCCGTA TGAACATCGGGCAAGTCCTAGAGTTACACTTGGGGATGGCTGCTCGCGAAATGGGCATCAAGAT TGCAACACCTGTCTTTGACGGTGCTAGTGAAGAAGATGTCTGGGAAACAGTTAAGGAAGCCGGC TTAGAAGCTGACGCTAAGACTATCTTATATGATGGTCGAACCGGTGAACCATTTGACCGTAAAG TCTCTGTTGGGGTTATGTACATGATTAAGTTGGCCCACATGGTCGATGACAAGTTGCACGCCCG STRR TTCAACAGGTCCATACTCTCTGGTTACCCAACAACCATTGGGTGGTAAAGCTCAATTTGGTGGG CAACGTTTCGGGGAGATGGAGGTTTGGGCCCTA -3'

SEQ ID n° 4: Partial sequence of the *rpoB* gene of *Streptococcus mutans*. This sequence measures 3198 base pairs, has a guanosine plus cytosine content of 42%, and is deposited with GenBank under number AF 535167.

TCAAGTCATTGGCTGAAAANNAGTAGATCTGAAACAGGCGAAATTCTTGTTGAAAGCTGGGACT GAAATGACACGCAGTGTAATTGATTCGATTGCAGATTATCTTGATGGAGATCTCAATAAAATTG TTTATACGCCAAATGAATACGCTGTTTTGACAGAACCTGTTGTTCTTCAAAAATTCAAAGTTAT GGCTCCAAATGATCCAGACCGCACGGTTACTGTTATTGGTAATGCCAGTCCAAGATGACAAAGT ACGTCACTTGACACCAGCCGATACGTATTAGCTGAAATGTCTTATTTCCTTAACTTGGCTGAGG GTNTAGGTAAAGTTGATGATATTGACCATTTAGGCAACCGACGTATTCGTGCTGTTGGTGAATT GCTTGCTAATCAATTTCGTATTGGTTTGGCACGTATGGAACGCAATGTTCGTGAACGCATGTCC GTTCAAGATAATGAAGTCTTAACGCCACAACAGATTATTAACATTCGCCCTGTAACAGCGGCAA TTAAAGAGTTTTTTGGTTCTTCTCAATTGTCACAGTTCATGGACCAACACACTCCACTGTCTGA ATTGTCTCATAAACGCCGTTTGTCAGCTTTAGGTCCTGGTGGTTTAACACGCGACCGTGCTGGT TATGAAGTCCGTGATGTGCACTATACGCÄTTATGGTCGTATGTGTCCAATTGAAACGCCTGAAG CCAAACACCATACCGTAAAGTTGACCGTGAGACAGGTAAAGTAACCAATGAAATCGAATGGCTT ACTGCTGATGAAGAAGATGAATTCACTGTAGCTCAGGCTAACTCAAAACTCAATGAAGATGGAA STRF GCTTTGCTGAAGAAATCGTCATGGGACGTCATCAAGGGAATAACCAAGAGTTTCCAGCAAGTTC TGTTGAATATATGGATGTTTCTCCTAAGCAGGTAGTTGCGGTAGCGACAGCATGTATTCCTTTC CTTGAAAATGATGACTCCAACCGTGCCCTTATGGGAGCTAACATGCAGCGCCAAGCTGTGCCAT TGATTGATCCTAAAGCACCTTTTGTTGGAACTGGTATGGAATATCAAGCAGCCCATGATTCTGG AGCCGCTATTATCGCTCAACATAATGGGAAAGTGGTTTATTCCGATGCAGATAAGATTGAAGTT CGCCGTGAAGATGGCTCACTAGATGTTTATCATGTTACCAAATTCCGTCGTTCTAACTCTGGAA CTGCCTACAATCAACGTACTCTTGTTAGGGTAGGCGATAGTGTTGAGAAGGGGGGACTTTATTGC AGATGGTCCTTCTATGGAAAAGGGTGAGATGGCTCTTGGACAAAATCCAGTGGTTGCTTACATG TTTATACTTCTGTCCATTTAGAAGAATTTGAATCTGAAACTCGTGATACAAAGCTTGGACCTGA AGAAATTACGCGTGAAATCCCAAATGTTGGTGAAGATGCCCTGAAAGACCTTGATGAAATGGGA ATTATTCGCATTGGTGCTGAGGTTAAAGAAGGTGATATTCTAGTTGGTAAAGTGACTCCTAAAG GAGAAAAAGATCTTTCTGCAGAAGAACGCCTCTTGCATGCCATTTTTGGTGACAAATCACGTGA TTTACACGTGCTAATGGAGATGAACTTCAATCAGGTGTTAACATGCTGGTTCGTGTTTATATCG CTCAAAAACGTAAAATCAAGGTCGGAGATAAGATGGCCGGACGTCATGGTAACAAGGGTGTCGT TTCCCGTATTGTACCAGTGGAAGATATGCCATATCTTCCAGATGGAACACCTGTTGATATCATG CTTAATCCACTTGGGGTGCCATCACGGATGAACATTGGGCAAGTTATGGAACTCCATCTTGGTA TGGCTGCTCGTAATTTGGGCATTCATATTGCAACGCCTGTCTTTGACGGAGCAACTTCTGATGA TCTTTGGGAAACAGTAAAAGAGCCGGTATGGATTCTGATGCTAAAACTGTTCTTTATGATGGT CGCACAGGGGAGCCGTTTGATAATCGTGTATCAGTTGGTGTTATGTATATGATTAAACTTCACC STRR ACATGGTTGATGAYAACCATTTTGTCTATGCAMAGWTCAGTTGGCCCTTAKTCAAYGAWTAMTC AGASGARTTCCTGCTWGGTGTAAAGGCTNCAATTGTCTTTAGAGGTTAAGGCTGGTGAAATAAC  ${\tt GGTATGCTGGTATTGATGGCAATGGGCAAGTGAATANTCAACACCGGCCGTCTACANCGTGC-3'}$ 

SEQ ID n° 5: Partial sequence of the rpoB gene of  $Enterococcus\ faecalis$ . This sequence measures 3096 base pairs, has a guanosine plus cytosine content of 42%, and is deposited with GenBank under number AF 535175.

5'-GACCCTTATCAATTGGTTTTTAGATGAGGGACTTCGTGAAATGTTTGAAGACATTTTACCAATT
GATGATTTCCAAGGAAACTTATCCTTAGAATTTGTTGACTATGAATTTAAAAGAACCAAAGTACA
CAGTAGAAGAAGCCCGCGCACATGATGCCAACTATTCTGCGCCATTACATGTAACATTACGTTT
AACCAACCGTGAAACAGGTGAAATTAAATCCCAAGAAGTCTTCTTCGGCGATTTCCCATTAATG
ACAGAAATGGGTACCTTCATCATCAACGGGGCAGAACGTGTTATCGTTTCCCAATTAGTTCGTT
CTCCAGGTGTTTACTTCCATGGAAAAGTGGACAAAAACGGCAAAGAAGGTTTTTGGCTCAACAGT
CATTCCTAACCGTGGTGCATGGTTAGAAATGGAAACAGATGCGAAAGACATTTCTTATGTTCGG
ATTGACCGCACACGTAAAATTCCTTTAACTGTGTTAGTTCGTGCTTTAGGTTTCGGTTCAGATG
ATACCATCTTCGAAATTTTCGGCGACAGCGAAAGCTTACGCAACACAATTGAAAAAGATTTACA
CAAAAACGCAAGTGATTCTCGTACAGAAGAAGACGTTTGAACACATTTATGAACGTCTTCGCCCA
GGCGAACCAAAAACAGCAGATAGCTCACGTAGCTTGTTAACTTGCACGTTTCTTTGATCCAAAA
CGTTATGATTTGGCAAACGTTGGTCGCTACAAAGTTAACAAAAAATTAGACTTAAAAACACGTC
TATTAAACTTAACCTTAGCTGAAACGCTAGTTGATCCAGAAACTGGTGTAAAATCATTGTCGAAA

AAGGCACAGTTTTAACACACTACATCATGGAAACATTAAGGCRATACATTGACAAACGGCTTAA ACAGCGTAACTTACTATCCAAGTGAAGATGCGGTAGTAACTGAACCAATGACGATCCAAGTGAT TCAAGTTCTTTCACCAAAAGATCCTGAACGTATCGTAAATGTGATTGGTAACGGCTATCCAGAC GACAGCGTAAAAACAGTTCGTCCAGCAGATATCGTTGCTTCAATGAGCTACTTCTTCAACTTAA TGGAAGATATCGGTAATGTCGATGACATCGACCACTTAGGTAATCGTCGTATCCGTTCAGTAGG CGAATTATTACAAAACCAATTCCGTATTGGTTTAGCCCCGTATGGAACGTGTGGTTCGTGAAAGA AGGTGAGTTAACCCATAAACGTCGTCTATCAGCCTTAGGGCCTGGTGGTTTGACTCGTGATCGT GCCGGTTATGAAGTTCGTGACGTTCACTACTCTCACTATGGTCGTATGTGTCCAATTGAAACGC CTGAGGGACCAAATATCGGGTTGATCAATAGCTTATCTAGTTATGCGAAAGTGAATAAATTTGG TTTCATCGAAACGCCTTATCGCCGTGTTGATCGTGCGACAGGCCGTGTTACTGATCAAGTAGAT TACTTAACAGCAGACATCGAAGACCATTATATCGTAGCGCAAGCGAACTCACTTTTAAATGAAG ATGGCACATTTGCCAATGATGTTGTTATGGCGCGTCTACAAAGTGAAAACTTAGAAGTTGCCGT AGACAAAGTTGACTACATGGACGTTTCACCAAAACAAGTAGTCGCAGTCGCAACAGCATGTATT CCTTTCTTAGAAAACGATGACTCCAACCGTGCCTTGATGGGTGCCAACATGCAGCGTCAAGCGG TGCCGTTAATTCAACCACGCTCTCCGTGGGTAGGTACAGGTATGGAATATAAATCAGCCCATGA CTCAGGTGCTGCTTTACTATGTAAACATGACGGTGTCGTAGAATTCGTCGATGCAAAAGAAATT STRF CGCGTTCGTCGCGACAATGGCGCATTAGACAAATATATGGTTACAAAATTCCGTCGTTCTAACT CAGGAACAAGCTACAACGCCCAATTGTTCACTTAGGTGAAAAGTTGAAAAGGCGATACTT TACCGGATGGACCTTCTATGGAAGAAGCGAAATGGCTTTATGGCAAAACGTCTTAGTTGCCTTC ATGACATGGGAAGGTTACAACTACGAGGATGCCATTATCATGAGCCGTCGTTTAGTTAAAGACG ATGTCTACACTTCTGTGCATATTGAAGAATATGAATCAGAAGCACGTGATACAAAATTAGGACC TGAAGAAATTACCCGTGAAATTCCAAACGTTGGGGAAGACGCGTTGAAAGACTTAGACGAAATG GGGATTATCCGCATTGGTGCTGAAGTTCAAGATGGCGACTTACTAGTTGGGAAAGTCACACCTA AAGGGGTCACAGAATTATCTGCAGAAGAACGTTTATTACACGCAATCTTCGGGGAAAAAGCCCG CGAAGTTCGTGATACGTCTCTCCGTGTACCTCACGGTGGCGGCGGTATCGTTCATGATGTGAAA ATCTTTACTCGTGAAGCTGGCGATGAATTATCACCAGGTGTCAACATGTTAGTTCGTGTCTATA TCGTTCAAAAACGTAAAATTCACGAAGGAGATAAAATGGCGGGACGTCACGGAAATAAAGGGGT TGTTTCCCGTATTATGCCGGAAGAAGATATGCCATTCTTACCTGACGGAACACCTGTTGATATC ATGTTGAACCCATTAGGGGTACCTTCTCGTATGAATATCGGACAAGTACTTGAATTACACTTAG GTATGGCTGCTCGCCAATTAGGTATTCACGTCGCAACACCTGTTTTCGATGGGGCCAACCGATGA AGACGTTTGGGAAACTGTTCGTGAAGCTGGTATGGCTAGCGATGCTAAAACAGTTCTTTACGAT GGACGTACAGGTGAACCATTTGATAACCGTATTTCCGTTGGTGTCATGTATATGATTAAATTAG CCCACATGGTTGATGACAAATTGCATGCTCGTTCAATCGGACCTTACTCTCTTGTTACGCAACA STRR ACCGTTGGGTGTAAAGCTCAATTC-3'

In the preceding sequences, the K nucleotide designates T or G, the M nucleotide designates A or C, the R nucleotide designates A or G, the W nucleotide designates A or T, the Y nucleotide designates C or T and the N nucleotide designates A, T, C or G.

Example 2: Partial sequencing of the rpoB gene 28 species of genus Streptococcus and related genera.

From the alignment of the complete sequences of the rpoB gene in Streptococcus spp. and Abiotrophia defectiva example 1 and those known in GenBank (Streptococcus pneumoniae AE008542 and Streptococcus pyogenes AE006480) a set of primers was chosen for the amplification and sequencing of a 709 to

740 bp fragment of this gene in 28 type strains of these bacterial genera. The sequences of these primers were:

- SEQ ID n° 6: 5'- AARYTIGMCCTGAAGAAAT-3'

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- SEQ ID n° 7: 5'- TGIARTTTRTCATCAACCATGTG-3'

Sequence SEQ ID n° 7 was used as 3' primer and therefore represents the complementary reverse sequence of the direct strand represented in preceding sequences SEQ ID n° 1 to 5.

These primers are incorporated with the DNA extracted from the bacteria during PCR under the following conditions: denaturing at 95°C for 1 min followed by 35 cycles comprising a denaturing step at 94°C for 10 sec, a hybridisation step at 52°C for 10 sec and an elongation step at 72°C for 30 sec.

The amplified products are sequenced with the same primers SEQ ID  $n^{\circ}$  6 and SEQ ID  $n^{\circ}$  7 under the following conditions: denaturing at 95°C for 1 min followed by 30 cycles comprising a denaturing step at 95°C for 30 sec, a hybridisation step at 52°C for 30 sec and a hybridisation step at 62°C for 1 min. The sequencing products are analysed on a ABI PRISM 3100 sequencer.

The inventors determined the position of these two primers SEQ ID  $n^{\circ}$  6 and SEQ ID  $n^{\circ}$  7, so as to observe the following criteria:

- 1-sequence flanked by these two primers specific to the species of the bacterium. This condition is verified after alignment of the fragments of around 720 bp with all the sequences of the *rpoB* bacterial genes available in computerized data banks,
- 2- search for the shortest possible identification region to achieve the best possible increase in the sensitivity of molecular detection,
- 3-primer length of 18 to 22 bp,
- 4-sequence of primers showing a close melting temperature,

5- sequence of primers not enabling auto-hybridisation or complementarity

The obtained *rpoB* gene fragments of the bacterial species of genus *Streptococcus* and said related genera have approximately 720 (709 to 732) base pairs and their sequence is specific to each species of this genus therefore permitting molecular identification of the bacteria of the 28 species tested, i.e.:

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- 10 SEQ ID n° 8: partial sequence of the rpoB gene in Streptococcus suis CIP 1032  $17^{T}$  measuring 709 base pairs:
  - 5'-CGCGAAATTCCAAACGTTGGTGAAGATGCCCTTCGCAACTTGGACGAAA
    TGGGGATTATCCGTATTGGTGCCGAAGTTAAAGAGGGCGACATTCTTGTTGG
    TAAAGTCACACCAAAAGGTGAAAAAGATCTTTCTGCTGAAGAGCGTCTCTTGC
    ACGCAATCTTCGGTGACAAGTCACGTGAAGTACCTCTCTTCGTGTA
    CCTCACGGTGCCGATGGTGTCGTTCGTGATGTGAAAATCTTTACTCGTGCCAA
    CGGTGATGAATTGCAATCAGGTGTTAACATGTTGGTTCGTGTTTACATCGCTC
    AAAAACGTAAGATCAAGGTCGGAGATAAGATGGCCGGTCGTCACGGTAACAA
    GGGTGTCGTTTCACGTATTGTACCTGTTGAGGATATGCCATATCTTCCAGATG
    GAACACCAGTTGACATCATGTTGAACCCACTCGGGGTGCCATCACGTATGAAC
    ATCGGTCAGGTTATGGAACTTCACTTGGGTATGGCGGCTCGCAACTTGGGCA
    TCCATATCGCAACACCAGTTTTCGATGGTGCAAGTTCAGAAGACCTCTGGTCA
    ACTGTTAAAGAAGCAGGTATGGACTCAGATGCCAAGACCATTCTTTACGATGG
    ACGTACAGGTGAACCATTTTGACAACCGTGTATCTTTTACGATGA
    TCAAGCTTCACCACATGGTTGATGACA

SEQ ID n° 9: partial sequence of the rpoB gene in Streptococcus sanguinis CIP 55.128<sup>T</sup> measuring 725 base pairs:

SEQ ID  $n^{\circ}10$ : partial sequence of the rpoB gene in Streptococcus salivarius CIP  $102503^{\mathsf{T}}$  measuring 728 base pairs:

SEQ ID n°11: partial sequence of the rpoB Streptococcus pyogenes CIP 56.41 measuring 725 base pairs: 5'- TGTCATCAACCATGTGGTGAAGTTTGATCATATACATGACACCAACGGAT ACACGGTTGTCAAATGGTTCACCGGTGCGACCATCATAAAGGACCGTCTTAGC ATCGCTATCCATACCAGCTTCACGAACAGTGTCCCAAAGGTCTTCTGATGAAG CCCCGTCAAAGACAGGTGTTGCAATGTGAATACCAAGATTACGAGCAGCCATA CCAAGGTGAAGTTCCATAACCTGACCAATATTCATCCGTGATGGCACCCCAAG AGGGTTCAACATGATGTCAACTGGTGTTCCGTCTGGAAGGTATGGCATGTCT TCAACTGGTACAATACGTGAAACGACACCCTTGTTTCCGTGACGACCGGCCAT TTTATCTCCGACCTTGATTTTACGTTTTTGAGCGATGTAAACACGCACAAGCAT ATTAACACCTGATTGCAATTCATCGCCGTTAGCGCGTGTAAAGATTTTCACATC ACGAACGATACCATCACCACCGTGAGGGACACGAAGTGAGGTATCACGCACT TCACGCGATTTATCCCCAAAGATGGCGTGAAGTAAACGTTCTTCAGCAGAAAG GTCTTTTCACCTTTAGGTGTGACTTTACCTACTAAGATGTCGCCTTCTTTAAC CTCAGCACCGATACGGATAATGCCCATTTCGTCAAGGTCTTTGAGGGCTTCTT CACCAACATTTGGGATTTCCGAGTGATTCTTCAGGGCA - 3'

5 SEO ID n°12: partial sequence of the rpoBgene in Streptococcus pneumoniae CIP 102911 measuring 724 base pairs: 5' - CAACCATGTGGTGGAGTTTGATCATGTACATGACTCCGACAGAAAACACG GTTATCAAACGGTTCACCAGTACGTCCATCGTAAAGGATCGTTTTGGCATCGC TATCCATACCTGCTTCTTTAACAGTTGACCAAAGATCTTCAGAACTTGCTCCAT CAAAGACTGGTGTCGCGATGTGAATACCAAGAGTACGAGCTGCCATACCAAG GTGAAGCTCCATAACCTGACCGATATTCATACGTGATGGTACCCCAAGTGGGT TCAACATGATGTCGACTGGAGTTCCGTCTGGAAGGTAAGGCATGTCTTCTACA GGAACGATACGAGAGACAACCCCTTTGTTTCCGTGACGTCCGGCCATTTTATC TCCGACCTTAATCTTACGTTTTTGAGCGATGTAAACACGAACCAACATGTTAAC ACCTGATTGCAACTCATCTCCATTTACACGTGTAAAGATCTTAACATCACGAAC GACACCATCGGCACCGTGTGGTACACGAAGAGAAGTATCACGCACTTCACGA GACTTGTCTCCAAAGATAGCGTGCAAGAGACGTTCTTCAGCTGAAAGATCTTT CTCACCCTTAGGTGTTACTTTACCTACAAGAATATCACCTTCTTTAACCTCAGCA CCAATACGGATAATCCCATTTCGTCAAGGTCTTTGAGGGCATCTTCACCAACG TTTTGGAATTTCGCGAGTGATTTCTTCAGGTCCAA - 3'

SEQ ID  $n^{\circ}13$ : partial sequence of the rpoB gene in  $Streptococcus\ oralis\ CIP\ 102922^{T}$  measuring 694 base pairs: 5'-

ACTCGTGAAATTCCAAACGTTGGTGAAGATGCCCTTAAAGACCTTGACGAAAT
GGGTATTATCCGTATTGGTGCTGAGGTTAAAGAAGGAGATATCCTTGTAGGT
AAAGTCACACCTAAGGGTGAAAAAAGACCTTTCTGCTGAAGAACGTCTCTTGCA
CGCTATCTTCGGAGACAAGTCTCGTGAAGTGCGTGATACTTCTCTTCGAGTAC
CTCACGGTGCCGATGGTGTCGTTCGTGATGTTAAGATCTTTACACGTGCAAAT
GGTGATGAGTTGCAATCTGGTGTGAATATGCTGGTTCGTGTCTACATCGCTCA
AAAACGTAAGATCAAGTCGGAGATAAGATGGCCGGACGTCACGGAAACAAAG
GGGTTGTCTCTCGTATCGTTCCTGTAGAAGACATGCCTTACCTTCCAGATGGA
ACTCCAGTCGATATCATGTTGAACCCACTTGGGGTGCCATCACGTATGAATAT
CGGTCAGGTTATGGAACTCCACCTTGGTATGGCAGCCCGTACTCTTGGTATCC
ACATCGCAACACCAGTCTTTGACGGAGCAAGTTCGGAAGACCTTTACGATGGAC
GTTAAAGAAGCAGGTATGGATAGCGATGCCAAAACAATCCTTTACGATGGAC
GTACAGGTGAGCCGTTTGACAACCGTGTATCAGTTGGTGTCATGTACATGATC
AAACTCCA—3'

SEO ID n°14: 5 partial sequence of the rpoBin Streptococcus mutans CIP 103220 measuring 728 base pairs: 5' - TGTCATCAACCATGTGGTGAAGTTTAATCATATACATAACACCAACTGATA CACGATTATCAAACGGCTCCCTGTGCGACCATCATAAAGAACAGTTTTAGCA TCAGAATCCATACCGGCTTCTTTTACTGTTTCCCAAAGATCATCAGAAGTTGCT CCGTCAAAGACAGGCGTTGCAATATGAATGCCCAAATTACGAGCAGCCATACC AAGATGGAGTTCCATAACTTGCCCAATGTTCATCCGTGATGGCACCCCAAGTG GATTAAGCATGATATCAACAGGTGTTCCATCTGGAAGATATGGCATATCTTCC ACTGGTACAATACGGGAAACGACACCCTTGTTACCATGACGTCCGGCCATCTT ATCTCCGACCTTGATTTTACGTTTTTGAGCGATATAAACACGAACCAGCATGTT AACACCTGATTGAAGTTCATCTCCATTAGCACGTGTAAAGATTTTCACATCACA AACAACACCGTCGCCACCATGAGGTACACGAAGAGAAGTATCACGAACTTCAC GTGATTTGTCACCAAAAATGGCATGCAAGAGGCGTTCTTCTGCAGAAAGATCT TTTTCTCCTTTAGGAGTCACTTTACCAACTAGAATATCACCTECTTTAACCTCAG CACCAATGCGAATAATTCCCATTTCATCAAGGTCTTTCAGGGCATCTTCACCAA CATTTGGGATTTCACGCGTAATTTCTTCAGGTCCA - 3'

SEQ ID  $n^{\circ}15$ : partial sequence of the rpoB gene in  $Streptococcus\ mitis\ CIP\ 103335^{T}\ measuring\ 730\ base\ pairs:$ 

5'-TGTCATCAACCATGTGGTGGAGTTTGATCATGTAACATGACTCCGACAGA
AAACACGGTTATCAAATGGTTCACCTGTACGTCCATCGTAAAGGATTGTTTTG
GCATCGCTATCCATACCAGCTTCTTTAACAGTTGACCAAAGATCTTCAGAACTT
GCTCCGTCAAAGACTGGTGTTGCGATGTGAATACCAAGAGTACGAGCTGCCA
TCCCAAGGTGGAGTTCCATAACCTGACCGATATTCATACGTGATGGCACCCCA
AGTGGGTTCAACATGATATCGACTGGAGTTCCATCTGGAAGGTAAGGCATAT
CTTCTACAGGAACGATACGAGAGACAACCCCTTTATTTCCGTGACGTCCGGCC
ATCTTATCTCCGACCTTGATCTTACGTTTTTGAGCGATGTAGACGCGAACCAG
CATGTTGACACCTGATTGCAATTCATCTCCATTTGCACGTGTAAAGATCTTAAC
ATCACGAACCACACCATCAGCTCCGTGTGGTACACGAAGAGAAGTGTCACGTA
CTTCACGAGATTTATCTCCGAAGATAGCGTGCAAGAGCCGTTCTTCAGCTGAA
AGGTCTTTCTCACCCTTAGGTGTTACTTTACCTACAAGGATATCCCCTTCTTTA
ACCTCAGCACCGATACGGATAATACCCATTTCGTCAAGATCTTTAAGGGCATC
TTCCCCAACGTTTGGGATTTCACGAGGTAATTTCTTCAGGTCCA - 3'

5 SEQ ID n°16: partial sequence of the rpoB gene in  $Streptococcus\ equinus\ CIP\ 102504^{T}$  measuring 697 base pairs: 5'-

SEQ ID n°17: partial sequence of the rpoB gene in Streptococcus constellatus CIP 103247<sup>T</sup> measuring 731 base pairs:

5'- AGTTGTCATCAACCATGTGTGCAATTTAATCATATACATGACACCGACAGA
TACACGGTTGTCAAACGGCTCGCCCGTACGACCATCATAAAGAATCGTCTTGG
CATCGCTATCCATGCCTGCTTCACGAACAGTATCCCAAAGGTCATCTGAGCTT
GCTCCGTCAAATACTGGCGTTGCTATGTGGATACCAAGGTTGCGAGCAGCCA
TACCAAGGTGAAGCTCCATAACCTGTCCGATATTCATACGTGATGGCACCCCA
AGTGGGTTCAACATGATGTCTACTGGTGTTCCGTCTGGAAGATAAGGCATAT
CCTCAACTGGAACGATACGGGAAACAACCCCTTTATTTCCGTGGCGTCCGGCC
ATCTTATCCCCAACGCGGATCTTTCGTTTTTTGAGCAATGTAAACACGCACCAAC
ATGTTGACACCAGATTGCAATTCATCACCGTTCGCACGAGTAAAGATTTTCAC
ATCACGGACAACCCCAGCACCACCATGTGGTACACGAAGAGATGTCTCACGTA
CTTCACGAGATTTATCACCGAAAAATTGCATGAAGCAGGCGTTCTTCAGCGGAT
AAGTCTTTTTCACCTTTCGGCGTTACTTTACCGACAAGAATGTCGCCCTCTTTC
ACCTCAGCACCAATGCGGATAATTCCCATTTCGTCAAGGTCTCTTAGCGCATCT
TCCCCAACGTTTGGAATTTCGCGCGTTAATTTCTTCAGGTCCAA - 3'

SEQ ID n°18: partial sequence of the rpoB gene in  $Streptococcus\ anginosus\ CIP\ 102921^{T}$  measuring 697 base pairs: 5'-

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CACGCGCGAAATTCCAAACGTCGGTGAAGATGCTTTGAGAGACCTTGACGAA
ACGGGAATTATCCGCATTGGTGCTGAGGTAAAAGAAGGCGACATTCTTGTCG
GTAAAGTAACACCGAAAGGTGAAAAAAGACTTATCTGCTGAAGAACGCCTGCT
TCATGCAATTTTCGGTGATAAATCTCGTGAAGTACGTGATACTTCCCTTCGTGT
ACCACATGGTGGTGCAGGGGTTGTCCGTGATGTGAAAATCTTTACTCGTGCG
AACGGTGATGAATTGCAATCTGGTGTCAACATGTTGGTACGTGTTTACATCGC
TCAAAAAACGGAAAATCCGTGTTGGGGATAAGATGGCTGGACGTCACGGAAAC
AAAGGGGTTGTTTCCCGCATTGTTCCAGTTGAGGATATGCCGTATCTTCCAGA
TGGAACACCAGTTGATATTATGTTGAACCCACTTGGGGTGCCATCTCGTATGA
ATATTGGTCAAGTTATGGAGCTTCACCTCGGTATGGCTCAGATGATCTTTGGGA
AACCGTTCGTGAAGCTGGCATGGATGCTCAGATGATCTTTTGGGA
AACCGTTCGTGAAGCTGGCATGGATAGCGATGCTCAGATCTTTTTGGAT
GGCCGTACTGGTGAGCCATTTGATAATCGTGTATCCGTTGGTGTCATGTACAT
GATCAAACTCCAC—3'

SEQ ID n°19: partial sequence of the rpoB gene in Streptococcus dysgalactiae CIP  $102914^{\rm T}$  measuring 728 base pairs:

5'-TGTCATCAACCATGTGGTGGAGTTTAATCATGTACATGACACCAACGGAT
ACACGGTTGTCAAATGGTTCGCCAGTACGTCCATCATAAAGGACCGTCTTAGC
ATCGCTATCCATACCAGCTTCACGAACAGTGTCCCAAAGGTCTTCTGATGAAG
CCCCGTCAAAGACAGGTGTTGCAATGTGAATACCAAGATTACGAGCAGCCATA
CCAAGGTGAAGTTCCATAACCTGACCAATGTTCATCCGTGATGGCACCCCAAG
AGGGTTCAACATGATGTCAACTGGTGTTCCATCTGGAAGGTATGGCATCTT
CAACTGGTACAATACGTGAAACGACACCCTTGTTTCCGTGACGACCAGCCATT
TTATCTCCGACTTTGATCTTACGTTTTTGAGCAATGTAAACACGCACAAGCATA
TTAACACCTGATTGCAATTCATCGCCGTTAGCGCGTGTAAAGATTTTCACATCA
CGAACGATACCATCACCACCGTGAGGTACACGAAGGGACGTATCACGAACTTC
ACGTGATTTATCTCCAAAGATGGCATGCAAGAGACGCTCTTCAGCAGAAAGGT
CTTTTTCACCTTTAGGTGTGACTTTACCTACTAAGATGTCGCCTTCTTTAACCTC
AGCACCGATACGGATAATTCCCATTTCGTCAAGGTCTTTGAGCGCTTCTTCACC
AACGTTTGGAATTTCGCGGGTGATTTCTTCAGGTCAA - 3'

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SEQ ID n°20: partial sequence of the rpoBgene in Streptococcus bovis CIP 102302 measuring 728 base pairs: 5' – TGTCATCAACCATGTGGTGAAGTTTGATCATGTACATGATACCAACAGAG ACACGATTATCAAATGGTTCACCTGTACGACCGTCATAAAGAACTGTCTTAGC GTCGCTATCCATACCAGCTTCACGAACAGTATCCCAAAGGTCTTCTGAAGTTG CCCCGTCAAAGACTGGAGTTGCAATGTGAATACCGAGGTTACGAGCTGCCAT ACCAAGGTGAAGTTCCATAACTTGTCCGATATTCATACGAGATGGCACCCCAA GAGGGTTCAACATGATATCAACTGGAGTTCCGTCTGGAAGATATGGCATGTC TTCAACAGGAACGATACGAGAAACAACCCCTTTGTTTCCGTGACGACCGGCCA TGTTGACACCTGATTGCAATTCATCACCGTTAGCACGTGTGAAGATTTTAACA TCACGAACACACCGTCTCCACCGTGTGGCACACGAAGTGATGTATCACGTAC TTCACGAGATTTATCACCGAAGATTGCGTGAAGAAGGCGTTCTTCAGCAGAAA GGTCTTTTCACCTTTAGGTGTTACTTTACCTACAAGGATATCACCTTCTTTAA CTTCAGCACCGATACGGATAATACCCATTTCGTCAAGGTCTTTAAGAGCTTCTT CACCAACGTTTGGAATTTCGCGAGTGATTTCTTCAGGTCAA - 3'

SEQID n°21: partial sequence of the rpoB gene in Streptococcus acidominimus CIP 82.4 measuring 728 base pairs:

CACGTGATTTGTCACCGAAGATAGCATGCAAGAGACGCTCCTCAGCAGAAAG
ATCTTTTTCACCTTTTGGTGTCACCTTACCAACAAGAATATCGCCTTCTTTAACT
TCTGCACCGATACGGATAATACCCCATTTCGTCAAGGTCTTTGAGGGCTTCTTC
ACCAACGTTTGGAATTTCACGAGTAATTTCTTCAGGTCA - 3'

SEQ ID n°22: partial sequence of the rpoB

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Streptococcus agalactiae CIP 103227<sup>T</sup> measuring 733 base pairs:
5'-TGAGTTGTCATCAACCATGTGGTGAAGTTTGATCATGTACATGACACCAA
CTGACACACGGTTATCGAATGGTTCACCAGTACGACCATCATAAAGAACAGTC
TTAGCATCTGAATCCATACCTGCTTCTTGAACAGTTTCCCAAAGGTCTTCTGAA
GAAGCCCCATCAAAGACTGGCGTTGCAATATGAATACCTAAATTACGAGCAGC

gene

in

SEQ ID n°23: partial sequence of the rpoB Streptococcus difficilis CIP 103768 measuring 714 base pairs: 5'- TTGTCATCAACCATGTGGTGAAGTTTGATCATGTACATGACACCAACTGAC ACACGGTTATCGAATGGTTCACCAGTATGACCATCATAAAGAACAGTCTTAGCAT CTGAATCCATACCTGCTTCTTGAACAGTTTCCCAAAGGTCTTCTGAAGAAGCCCC ATCAAAGACTGGCGTTGCAATATGAATACCTAAATTACGAGCAGCCATACCTAAA TGAAGCTCCATAACTTGTCCGATATTCATACGTGATGGCACCCCAAGTGGGTTCA ACATGATATCAACTGGCGTTCCATCTGGTAAATAAGGCATATCTTCAACAGGAAC AATACGTGAGACGACACCTTTGTTTCCGTGACGACCGGCCATCTTATCACCGACT TTGATTTTACGTTTTTGAGCGATATAAACGCGGACAAGCATATTAACACCTGATT GCAATTCATCACCATTTGCACGAGTAAAGATTTTAACGTCACGAACTACTCCATC GCCACCGTGAGGTACACGTAGTGAAGTATCACGAACTTCACGTGATTTATCACCA AAAATGGCATGCAAGAGACGTTCTTCAGCAGATAAGTCCTTTTCACCCTTAGGCG TTACCTTACCAACAAGAATGTCACCTTCTTTTACCTCAGCACCAATGCGGATAATT CCCATTTCATCGAGATCACGTAGTGAATCTTCACCAACATTTGGAATTTCACGAG TA - 3

5 SEQ ID n°24: partial sequence of the rpoB gene in Streptococcus intermedius CIP  $103248^{\rm T}$  measuring 728 base pairs:

5'-TGTCATCAACCATGTGGTGAAGCTTAATCATGTACATGACACCAACGGAC
ACACGGTTATCAAACGGTTCGCCAGTACGTCCATCATAAAGGATTGTCTTAGC
ATCGCTATCCATACCTGCTTCACGAACGGTTTCCCAAAGATCATCTGAGCTAGC
TCCGTCAAAGACTGGCGTTGCAATGTGGATACCAAGTTGCGAGCAGCCATAC
CGAGGTGCAATTCCATAACTTGTCCGATATTCATACGTGACGGCACCCCAAGA
GGATTCAACATGATATCAACTGGTGTCCCGTCTGGAAGATACGGCATATCCTC
AACTGGAACAATGCGGGAAACAACCCČTTTGTTTCCGTGGCGTCCGGCCATCT
TATCTCCAACGCGGATTTTCCGTTTTTTGAGCGATATAAACACGTACCAACATGT
TGACACCGGATTGCAATTCATCACCGTTCGCACGAGTAAAGATTTTTACATCAC
GGACAACACCTGCACCACCGTGTGGTACACGAAGGGGAGGTATCACGCACTTC
ACGAGACTTATCACCAAAAAATTGCATGAAGCAGGCGTTCTTCAGCGGATAAAAT
CTTTTTCACCTTTCGGCGTTACTTTACCGACAAGAATGTCGCCTTCTTTTACCTC
AGCACCAATGCGGATAATTCCCATCTCGTCAAGGTCTCTCAAAGCATCTTCCCC
GACGTTTGGAATTTCGCGCGTTGATTTCTTCAGGTCCA-3'

SEQ ID  $n^225$ : partial sequence of the rpoB gene in  $Streptococcus\ equi\ CIP\ 102910^T$  measuring 728 base pairs:

5 SEQ ID n°26: partial sequence of the rpoB gene in  $Enterococcus\ gallinarum\ CIP\ 103013^T$  measuring 694 base pairs: 5'-

SEQ ID  $n^2$ 7: partial sequence of the rpoB gene in Enterococcus casseliflavus CIP  $103018^T$  measuring 727 base pairs:

5'-TGTCATCAACCATGTGGGCCAATTTGATCATGTACATGACACCAACGGAG
ATGCGGCCATCAAATGGTTCGCCGGTACGTCCGTCGTAAAGCACTGTTTTGGC
ATCGCTGGCCATTCCTGCTTCAGCAACCGTTGCCCAAACATCTTCATCGCTGGC
TCCATCAAAGACTGGTGTTGCCACGTGAATGCCTAATTGACGCGCAGCCATTC
CTAAGTGTAACTCTAATACTTGTCCAATGTTCATCCGAGAAGGTACCCCTAATG
GGTTCAGCATGATATCGACTGGTGTGCCATCTGGTAAGAAAGGCATGTCTTCT
TCTGGCATAATGCGAGAAACGACCCCTTTGTTTCCGTGACGTCCGGCCATTTT
ATCCCCTTCATGGATTTTCCGTTTTTGAACGATATAAACGCGAACCAGCATGTT
CACACCTGGTGACAATTCATCGCCAGCTTCGCGGGGTAAAGATTTTGACATCGT
GGACGATTCCGCCGCCGCCGTGAGGCACGCGTAGAGAAGTTTCACGCACTTC
GCGGGCTTTTTCACCAAAAGATTGCGTGCAACAAACGCTCTTCTGCTGAAAGTT
CCGTTACCCCTTTTGGCGTGACTTTCCCCAACAAGCAGTCTTTCACCC
AACGTTCGGGGATTTCGCGAGTGATTTCTTCAGGTCTTTCAACGCGTCTTCCC
AACGTTCGGGGATTTTCGCGAGTGATTTCTTCAGGTCCAA-3'

SEQ ID n°28: partial sequence of the rpoB gene in Enterococcus saccharolyticus CIP  $103246^{T}$  measuring 721 base pairs:

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5'-TGTCATCAACCATGTGGGCAAGTTTAATCATGTACATTACCCCAACAGAG
ATACGACCATCGAATGGTTCACCCGTACGTCCGTCATAAAGAACAGTTTTCGC
ATCGCGCGCCATGCCCGCTTCGCGAACTGTTTCCCATACGTCATCATCTGATGC
ACCATCAAATACTGGTGTAGCTACATGGATGCCTAACTGACGTGCAGCCATCC
CTAAGTGTAATTCCAATACTTGTCCGATGTTCATACGAGATGGTACTCCTAGT
GGGTTCAACATGATATCAACTGGTGTGCCGTCTGGTAAGAATGGCATGTCTTC
TTCTGGCATAATGCGAGAGACAACCCCTTTGTTACCATGACGTCCCGCCATTTT
ATCTCCTTCGTGAATCTTACGTTTTTGCACGATATAAACACGAACTAACATGTT
CACACCTGGAGATAATTCGTCGCCTGCTTCACGGGTAAAGATTTTAACATCGT
GAACGATACCGCCACCGCCGTGAGGAACACCGTAATGATGTATCACGTACTTCA
CGTGCTTTTTCACCGAAGATTGCGTGCAATAGACGTTCTTCTGCAGATAATTC
GGTTACCCCTTTAGGAGTGACTTTACCTACTAATAAGTCGCCATCTTGTACTTC
GGCACCGATACGGATAATACCCATTTCGTCTAAGTCTTTTAATGCGTCTTTCCCC
AACGTTAGGAATTTCGCGTGTATTCTTCAG - 3'

SEQ ID n°29: partial sequence of the rpoB in Enterococcus faecium CIP  $103014^{T}$  measuring 727 base pairs: 5'- TGTCATCAACCATGTGAGCAAGTTTGATCATGTACATCACACCGACAGAC ACACGTCCATCAAATGGTTCACCTGTACGTCCGTCGTACAGAACAGTTTTCGC ATCGCTGGCCATACCGGCTTCACGAACTGTTTCCCATACGTCTTCATCACTTGC ACCATCAAATACTGGCGTTGCTACGTGGATACCTAACTGACGTGCAGCCATAC CCAAGTGTAATTCCAATACTTGCCCGATGTTCATACGTGAAGGCACCCCTAAA GGATTCAGCATGATATCGATTGGTGTTCCATCAGGTAGGAATGGCATATCTTC TTCCGGCATAATACGGGATACAACCCCTTTATTTCCGTGACGACCGGCCATTTT ATCCCCTTCATGGATTTTACGTTTTTGAACGATATAAACACGAACTAACATGTT TACGCCTGGTGACAATTCATCTCCAGCTTCACGAGTAAAGATTTTCACATCGT GAACGATACCGCCGCCGCCATGTGGTACACGTAATGATGTATCGCGGACTTCA CGAGCTTTTTCGCCAAAGATCGCATGCAATAGACGTTCTTCTGCAGATAATTCT GTTACCCCTTTTGGCGTGACTTTCCCTACAAGCAAATCGCCATCTTGGACTTCT GCACCAATACGGATGATACCCATTTCGTCTAAATCTTTTAATGCGTCTTCCCGA CATTAGGGATTTCGCGTGTGATTTCTTCAGGTCCA - 3'

SEO ID n°30: partial sequence of the rpoBgene in Enterococcus faecalis CIP 103015 measuring 724 base pairs: 5'- TGTCATCAACCATGTGGGCTAATTTAATCATACATGACACCAACGGAA ATACGGTTATCAAATGGTTCACCTGTACGTCCATCGTAAAGAACTGTTTTAGC ATCGCTAGCCATACCAGCTTCACGAACAGTTTCCCAAACGTCTTCATCGGTTGC CCCATCGAAAACAGGTGTTGCGACGTGAATACCTAATTGGCGAGCAGCCATAC CTAAGTGTAATTCAAGTACTTGTCCGATATTCATACGAGAAGGTACCCCTAAT GGGTTCAACATGATATCAACAGGTGTTCCGTCAGGTAAGAATGGCATATCTTC TTCCGGCATAATACGGGAAACACCCCTTTATTTCCGTGACGTCCCGCCATTTT ATCTCCTTCGTGAATTTTACGTTTTTGAACGATATAGACACGAACTAACATGTT GACACCTGGTGATAATTCATCGCCAGCTTCACGAGTAAAGATTTTCACATCAT GAACGATACCGCCGCCACCGTGAGGTACACGGAGAGACGTATCACGAACTTC GCGGGCTTTTTCCCCGAAGATTGCGTGTAATAAACGTTCTTCTGCAGATAATT CTGTGACCCCTTTAGGTGTGACTTTCCCAACTAGTAAGTCGCCATCTTGAACTT CAGCACCAATGCGGATAATCCCCATTTCGTCTAAGTCTTTCAACGCGTCTTCCC AACGTTTGGAATTTCACGGGTATTTCTTCAGGTCA - 3'

SEQ ID n°31: partial sequence of the rpoB gene in  $Enterococcus\ avium\ CIP\ 103019^T$  measuring 570 base pairs:

5'- GTCCATCATAAAGAACGGTCTTAGCATCTGCTGCCATACGAGCTTCACGA
ACTGTTTCCCAAACATCGCTATCTTGCGCACCATCGAAGACTGGTGTCGCAAC
ATGGATACCTAGTTGGCGAGCCGCCATTCCCAAGTGTAATTCCAACACTTGTC
CGATGTTCATCCGAGATGGCACACCTAATGGGTTCAACATGATATCAACTGGC
GTACCGTCTGGTAAGAAAGGCATGTCTTCTTCTGGCATAATGCGAGAAACGA
CCCCTTTATTTCCGTGACGGCCGGGCATTTTATCCCCTTCATGAATCTTACGTT
TTTGCACGATGTACACGCGCACTAACATATTTACACCTGGAGATAATTCATCGC
CTGCTTCACGAGTAAAGATCTTCACATCGTGAACGATCCCGCCGCCACCATGC
GGTACACGAAGAGATGTATCACGAACTTCACGAGCCTTTTCACCAAAGATCGC
ATGCAACAAACGTTCTTCAGCTGATAATTCTGTTACCCCTTTAGGAGTGACTTT
ACCAACTAATAAATCACCATCATGAACTTCAGCACCAATAC -3'

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SEO ID n°32: partial sequence the of rpoB gene Abiotrophia defectiva CIP 103242<sup>T</sup> measuring 732 base pairs: 5'- GAAGTTGTCATCAACCATGTGGGCCAACTTAATCATGTACATAACCCCAA CAGAGACTTTACGGTCAAATGGTTCACCGGTTCGACCATCATATAAGATAGTC TTAGCGTCAGCTTCTAAGCCGGCTTCCTTAACTGTTTCCCAGACATCTTCTTCA CTAGCACCGTCAAAGACAGGTGTTGCAATCTTGATGCCCATTTCGCGAGCAGC CATCCCCAAGTGTAACTCTAGGACTTGCCCGATGTTCATACGGGATGGAACCC CTAATGGGTTCAACATGATATCAACTGGGGTACCATCTGGTAAGAATGGCATA TCTTCTTCCGGCATGATAAGGGAGACAACCCCTTTGTTACCGTGACGACCGGC CATCTTATCCCCTTCATTGATTTTACGTTTTTGTACGATGTAGACGCGGACTAG CTTGTTGACACCTGGTGCCAATTCGTCGCCAGCTTCGCGGGTAAAGATTTTAA CGTCGTGGACAATCCCGCCCCCCCGCCGTGTGGCACACGCAAGGAAGTATCACG TACTTCACGCGCCTTCTCACCGAAGATAGCATGGAGCAAGCGTTCTTCCGCAG ACAACTCGGTCACACCTTTTGGTGTTACCTTACCAACTAAGATATCGCCGTCTT TTACTTCCGCCCCGATACAGATAATCCCGTCTTGGTCTAAGTACTTGAGGGCA TCTTCGGACACGTTTGGAATTTCGCGTGTAATTTCTTCAGGTCA - 3'

SEQ ID n°33: partial sequence of the rpoB gene in Gemella morbilorum CIP  $81.10^{T}$  measuring 727 base pairs:

5'-TGTCATCAACCATGTGTGCAAGTTTATCATGTACATTACCCCTACAGATAC
ACGGCTATCAAATGGCTCACCTGTACGTCCGTCATAAAGAACTGTCTTAGCAT
CTTTAGCCATTCCAGCTTCCGCAACTGTAGACCAAACATCTTCATCAGTAGCAC
CATCGAATACTGGTGTAGCTACGTGGATTCCAAGTTGTTTAGCAGCCATACCT
AAGTGTAGCTCTAATACTTGTCCAATGTTCATACGAGATGGAACCCCAAGTGG
GTTTAACATTACGTCAACTGGTGTACCATCTGGTAGGTAAGGCATATCTTCTT
CTGGTAAGATATTTGAGATAACCCCTTTGTTACCGTGACGACCGGCCATTTTA
TCTCCTACACGAATTTTACGTTTTTTGGACGATAAATACACGAACAAGTTCATTT
ACACCGTTAGGTAATTCAGCACCATCTTCACGTTTAAAGATTTTAACATCAGCA
ACTACTCCATCAGCACCGTGAGGTACACGTAATGAAGTATCACGTACTTCTTTA
GATTTAGCTCCAAAGATAGCATATAATAATTTTTCTTCTGGAGTTTGTTCAGTT
AATCCTTTCGGTGTAACTTTACCTACTAAAATATCTCCCATCTTTAACTTCAGCC
CCAATACGAATGATTCCTCGTGCATCTAAGTTTCTAAGTGCATTTTCACCCTAC
GTTTGGAATCTCACGAGTAATTTCTTCAGGTCA - 3'

5 SEQ ID n°34: partial sequence of the rpoB gene in Gemella haemolysans CIP  $101126^{T}$  measuring 726 base pairs:

SEQ ID n°35: partial sequence of the rpoBGranulicatella adjacens CIP 103243<sup>T</sup> measuring 719 base pairs: 5'- CATCAACCATGTGAGCAAGTTTGATCATGTACATAACCCCTACTGACACA CGGTTATCGAATGGTTCCCCTGTACGTCCATCATATAGAATTGTTTTCGCATCA CGAGCCATACCCGCTTCTGCAACAGTTCCCCATACGTCTTCATCTTGCGCACCA TCGAATACTGGTGTTGCGATGTAAATACCTAATTCACGAGCAGCCATCCCTAA GTGTAACTCTAACACTTGTCCGATGTTCATACGTGAAGGTACCCCTAATGGGT TTAACATGATGTCAACTGGTGTTCCATCTGGTAAGAATGGCATATCTTCTTCC GGCATAATACGGGAAACAACCCCTTTATTACCGTGACGTCCGGCCATCTTATC CCCTTCATTGATTTTACGTTTTTGTACAATATATACACGAACTAATTTGTTTACG CCAGGTGCTAATTCATCACCTGCTGCACGTGTGAATACACGTACATCACGGAC AATACCGCCACCGCGTGAGGTACACGTAGAGATGTGTCACGAACTTCACGA GCTTTTCACCGAAGATTGCGTGTAATAAACGTTCCTCTGGTGATTGTTCTGTT AACCCTTTAGGAGTTACTTTACCAACTAAGATGTCACCATCTTTAACTTCGGCA CCGATACGAATAATTCCGTCTGCGTCTAGGTTCTTCAATGCGTCTTCCCAACGT TTGGAATCTCACGAGTAATTCTTCAGG-3°

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In the above sequences, the M nucleotide designates A or C, the R nucleotide designates A or G, the W nucleotide designates A or T, the Y nucleotide designates C or T and the N nucleotide designates A, T, C or G.

In the above sequences, the CIP references relate to deposits with the national collection of microorganism cultures: Collection Nationale de Culture des Microorganismes (CNCM) at Institut Pasteur in Paris (France).

15 Example 3: Blind identification of a collection of 20 bacterial strains comprising 10 strains of bacteria belonging to genus *Streptococcus* and related genera.

A collection of twenty strains belonging to the following bacterial species: Streptococcus pyogenes, Streptococcus sanguis, Granulicatella adjacens, Abiotrophia defectiva, Enterococcus avium, Enterococcus faecalis, Gemella

haemolysans, Gemella morbilorum, Streptococcus Streptococcus anginosus, Staphylococcus aureus, Pseudomonas Mycobacterium oleovorans, avium, Bacillus cereus, Acinetobacter anitratus, Corynebacterium amycolatum, 5 Klebsiella terrigena, Pasteurella, Lactobacillus rhamnosus, Staphylococcus was coded so as to conduct blind molecular identification of strains (the experimenter not having any a knowledge of strain identity) using the described in the present patent application. Extraction of the 10 nucleic acids and amplification of the rpoB gene fragment were performed as described in example 2 incorporating primers consisting of mixtures of 4 oligonucleotides which have sequences consisting of sequences SEQ ID n°6 (as 5' and SEQ ID n°7 (as 3' primer) where N represents inosine, in a 15 amplification (Fig.1). The sequencing of these 10 amplificates conducted was by incorporating into the sequencing reaction the primers SEQ ID n° 6 and SEQ ID n° 7 as described in example 2, and comparison of the sequences obtained with sequences SEQ ID n° 1 to 5 and 8 to 35 enabled 20 the 10 ten amplified strains to be identified as being Streptococcus pyogenes, Streptococcus sanguis, Granulicatella Abiotrophia defectiva, Enterococcus Enterococcus faecalis, *Gemella* haemolysans, Gemella morbilorum, Streptococcus equi, Streptococcus anginosus. 25 decoding of these 10 strains showed 100% agreement between molecular identification using the method that is the subject of the invention and the identification previously established by standard phenotype methods. This result illustrates the specificity of the set of primers SEQ ID n°6/SEQ ID n°7.

The other bacteria chosen because they are frequently isolated in human or animal clinical specimens and also possibly contain bacteria of genus *Streptococcus* were not amplified, thereby exhibiting the specificity of the primers

used for the *Streptococcus* genus and said 4 related genera under the conditions of use of the invention for detecting bacteria of genus *Streptococcus* and said 4 related genera in comparison with bacteria of another genus.

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Figure 1 shows the PCR amplification products obtained from ten coded bacterial strains, comprising 7 strains belonging to genus *Streptococcus* and said 4 related genera (columns 2,3,4, 7-11) and 3 bacterial strains of bacterial genera other than *Streptococcus* and said 4 related genera (columns 5, 6 and 12). Columns 1 and 13 show the molecular weight marker. The amplification products are obtained after incorporating primers SEQ ID n° 6 and SEQ ID n° 7 described above, and are visualized by staining with ethidium bromide after electrophoresis on agarose gel.